

Unraveling the Potential of Mesenchymal Stem Cell-Derived Extracellular Vesicles in Combating Neuroinflammation and Mitochondrial Dysfunction in Alzheimer's Disease

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Abstract

Alzheimer's disease (AD) stands as a formidable, age-related neurodegenerative disorder that is marked by the relentless degeneration of diverse neuronal cell types. This degeneration leads to profound losses in neural tissue, disruption of synaptic networks, cerebral atrophy, and a debilitating decline in cognitive function. Accordingly, the multifaceted nature of AD necessitates a robust and comprehensive therapeutic approach. Although current pharmacological treatments mainly focus on modulating acetylcholine levels through the inhibition of acetylcholinesterase, they only provide symptomatic relief and fail to alter the disease's relentless progression. In this context, mesenchymal stem cell-derived extracellular vesicles (MSC-EVs) emerge as a groundbreaking therapeutic strategy with remarkable potential. These vesicles can modulate neuroinflammation, influence immunological responses, orchestrate microglial activation, and reduce oxidative stress, thereby slowing AD progression. The inflammatory microenvironment plays a pivotal role in AD pathogenesis, with proinflammatory cytokines contributing to neurotoxicity and neuronal apoptosis. Moreover, due to their ability to cross the blood-brain barrier, MSC-EVs are considered key players in delivering therapeutic effects directly within the central nervous system. More precisely, they promote neural tissue regeneration, stimulate neurogenesis, enhance the reorganization of neuronal circuits, and improve synaptic plasticity, thereby fostering essential functional recovery. Therefore, this review highlights the transformative potential of MSCs-EVs in combating the progression of AD, presenting them as a vital pathway for the innovative treatment of neurodegenerative disorders.

Keywords: Alzheimer's disease, Pathophysiology, MSCs-EVs, Neuroinflammation, Mitochondrial conditions, Microglial activation

Introduction

Alzheimer's disease (AD) is regarded as the most prevalent neurodegenerative disorder, pathologically characterized by the extracellular aggregation of beta-amyloid (A β) plaques and the formation of intracellular neurofibrillary tangles (NFTs) composed of hyperphosphorylated tau protein.¹ In addition, this disorder is associated with synaptic degeneration, neuronal loss, and gliosis.² Clinically, AD frequently begins with mild cognitive impairment, manifesting as early memory deficits and difficulties with short-term memory retention.³ As the disease progresses, patients may exhibit increasingly severe impairments in complex attention, expressive language, visuospatial processing, and executive function. It should be noted that AD accounts for approximately 60–80% of all dementia cases.⁴ While around 1% of cases

are early-onset and exhibit an autosomal dominant inheritance pattern, typically manifesting symptoms before age 65, often in individuals' 40s or 50s, the vast majority (99%) are late-onset, sporadic cases.⁵

Pathophysiology of Alzheimer's Disease

AD pathophysiology is primarily driven by the accumulation of A β in the brain, which is linked to the amyloid precursor protein (APP) gene on chromosome 21.⁶ APP exists in three isoforms, with APP695 being predominantly found in neurons. Moreover, it can be processed through the amyloidogenic pathway, which produces neurotoxic A β peptides, and the non-amyloidogenic pathway, which prevents A β formation. In the amyloidogenic pathway, APP is cleaved by β -secretase (beta-site amyloid precursor protein cleaving enzyme-1:

BACE1) and γ -secretase, generating A β 40 and A β 42, with A β 42 being particularly harmful.⁷ Furthermore, the non-amyloidogenic pathway yields protective fragments that facilitate synaptic plasticity. The amyloid cascade hypothesis proposes that A β accumulation triggers the formation of NFTs, ultimately leading to cognitive decline. Tau, essential for microtubule stability and neuronal transport, becomes dysfunctional in AD, disrupting these functions and causing cognitive deficits.⁸ The pathological hyperphosphorylation of tau leads to the formation of NFTs, which begin in the entorhinal cortex and spread to other brain areas.⁹ Additionally, dysfunctions in lysosomal pathways promote tau accumulation and spread, while A β deposition accelerates tau pathology. Overall, A β and tau contribute to synaptic dysfunction and neuronal death (Figure 1).²

Genetic Background and Relevant Risk Factors

Genetic predisposition is a critical component in AD pathophysiology, accounting for approximately 58–79%

of cases. The autosomal dominant forms of AD are linked to rare mutations in the *APP*, *PSEN1*, and *PSEN2* genes.¹⁰ Among genetic risk factors, the apolipoprotein E (APOE) gene is the most significant for sporadic AD.¹¹ The prevalence of the APOE ϵ 4 allele is reported to be around 66% in individuals diagnosed with AD and approximately 64% among those with mild cognitive impairments. The presence of a single APOE ϵ 4 allele escalates the risk of developing AD by 3–4 times, while individuals carrying two alleles face a risk increase of 9–15 times.¹² Additionally, age remains the predominant risk factor, with an estimated prevalence of AD at 18.1% in individuals aged 65 and older, which surges to 33.2% for those aged 85 and above.¹³

Mitochondrial Dysfunction in Alzheimer's Disease Development

Mitochondrial abnormalities are increasingly recognized in the pathogenesis of neurodegenerative diseases, such as AD, Parkinson's, and Huntington's. Mitochondrial

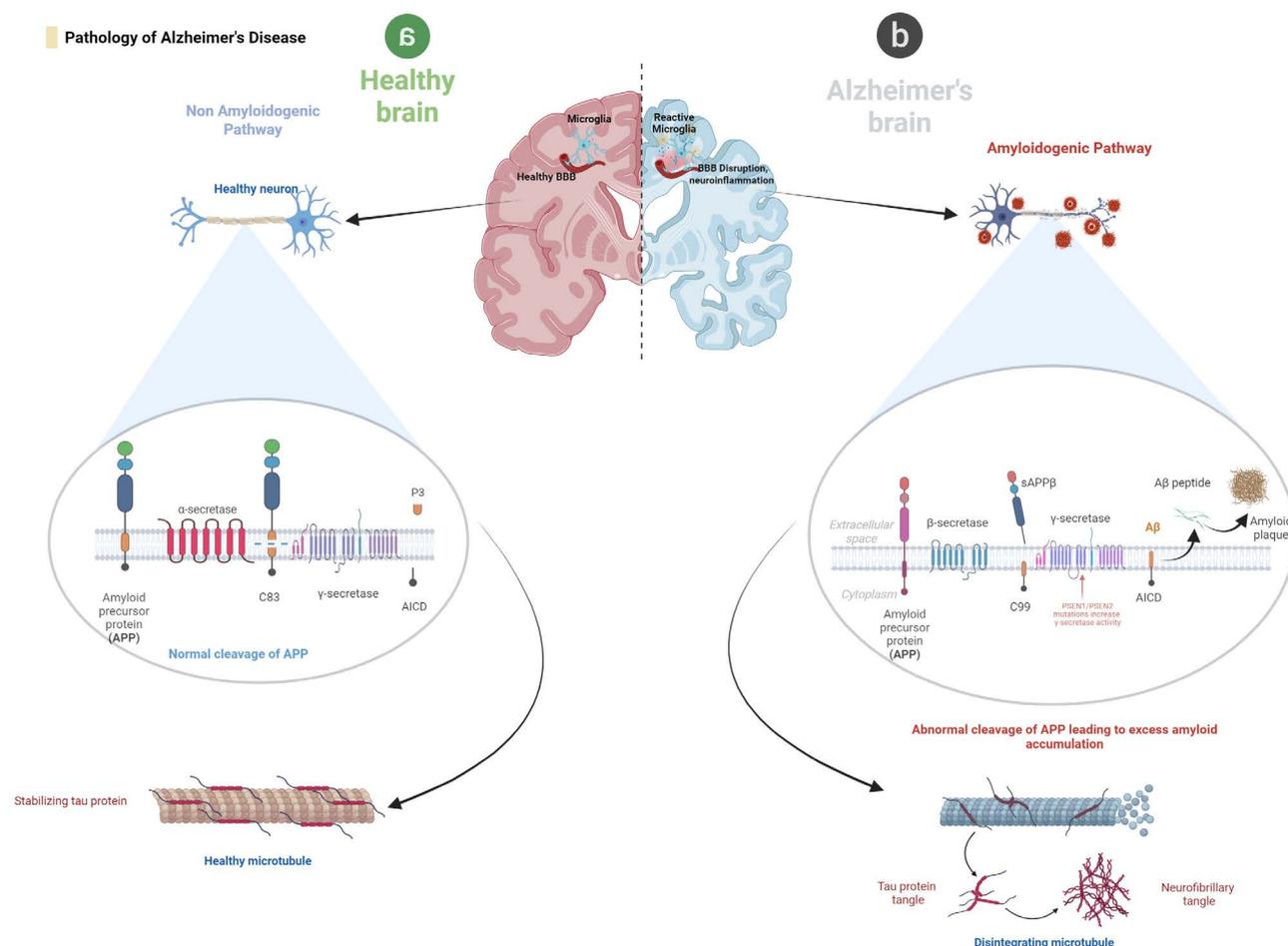


Figure 1. Pathophysiological Mechanisms Underlying Alzheimer's Disease

Note. APP: Amyloid-beta precursor protein; BBB: Blood–brain barrier; A β : Beta amyloid; AD: Alzheimer's disease. The left panel illustrates the healthy brain, in which the APP undergoes normal non-amyloidogenic cleavage by α -secretase and γ -secretase, generating non-pathogenic fragments and preventing formation. A β plays a major role in synaptic development, mineral transfer, neuronal plasticity, and regulation of cell signaling in neuronal tissues. Neurons retain stable microtubules supported by physiologic tau activity, and both the BBB and microglial populations remain in a resting, protective state. The right panel displays the amyloidogenic pathway of APP processing, which characterizes the general pathology of AD. The sequential cleavage of APP by β -secretase and γ -secretase produces A β peptides that aggregate and accumulate into amyloid plaques. Abnormal tau modification leads to tau tangles and microtubule disassembly, resulting in the instability of microtubules, which play a crucial role in neuronal cell structure. It should be noted that such molecular events promote neuroinflammation, reactive microglia, and BBB disruption characteristics of AD progression.

dysfunction, metabolic issues, increased reactive oxygen species (ROS), and impaired mitochondrial quality control (QC) are early indicators in AD and play a crucial role in disease progression.¹⁴ In addition, genome-wide studies have linked variants in the *ABCA7* gene to late-onset AD. Research using cortical organoids derived from *ABCA7*-deficient-induced pluripotent stem cells (iPSCs) demonstrates serious neuronal damage, including disrupted mitochondrial lipid metabolism and excessive ROS production, both of which are critical for cognitive function. Mitochondrial QC involves dynamic processes, such as fission, fusion, and mitophagy, and impairments in these processes may contribute to neurodegenerative disorders (Figure 2).¹⁵ Furthermore, mitochondrial fission and fusion processes are integral to mitochondrial dynamics and cellular homeostasis. Fission, the process of mitochondrial division, enables mitochondrial fragmentation, thereby facilitating the removal of damaged components and allowing for the effective distribution of mitochondria during cell division. Conversely, fusion involves the merging of mitochondria, promoting the mixing of mitochondrial contents and DNA, which is essential for maintaining healthy, functional organelles. In general, these dynamic processes are critical for regulating mitochondrial function, influencing adenosine triphosphate (ATP) production, metabolic efficiency,

and the cellular stress response, and ultimately impacting overall cellular health and metabolic homeostasis (Figure 3).¹⁶ Early Alzheimer's models show reduced mitophagy in iPSC-derived neurons. The accumulation of APP cleavage products at mitochondria can lead to mitophagy deficits. In familial Alzheimer's, iPSC-derived neurons frequently have defective mitochondria due to compromised autophagic degradation linked to lysosomal dysfunction.¹⁷ Moreover, mutant PS1 M146L iPSC-derived cells exhibit defects in respiratory chain enzymes and autophagy-related proteins, influencing AD pathology. Further, both nuclear and mitochondrial DNA mutations play critical roles in the development of the disease and the aging process.¹⁸

Oxidative Stress and Alzheimer's Disease Development

An imbalance in ROS production and accumulation causes OS. ROS are essential signaling molecules in biological systems, but excess ROS can harm cells.^{19,20} They include chemically reactive oxygen-free radicals and oxygen derivatives that are not radicals. A cell might enter an oxidative state if it produces more ROS or if its antioxidant system is not functioning well. The brain is susceptible to oxidative damage due to excessive oxygen consumption, elevated concentrations of polyunsaturated fatty acids, increased levels of redox-

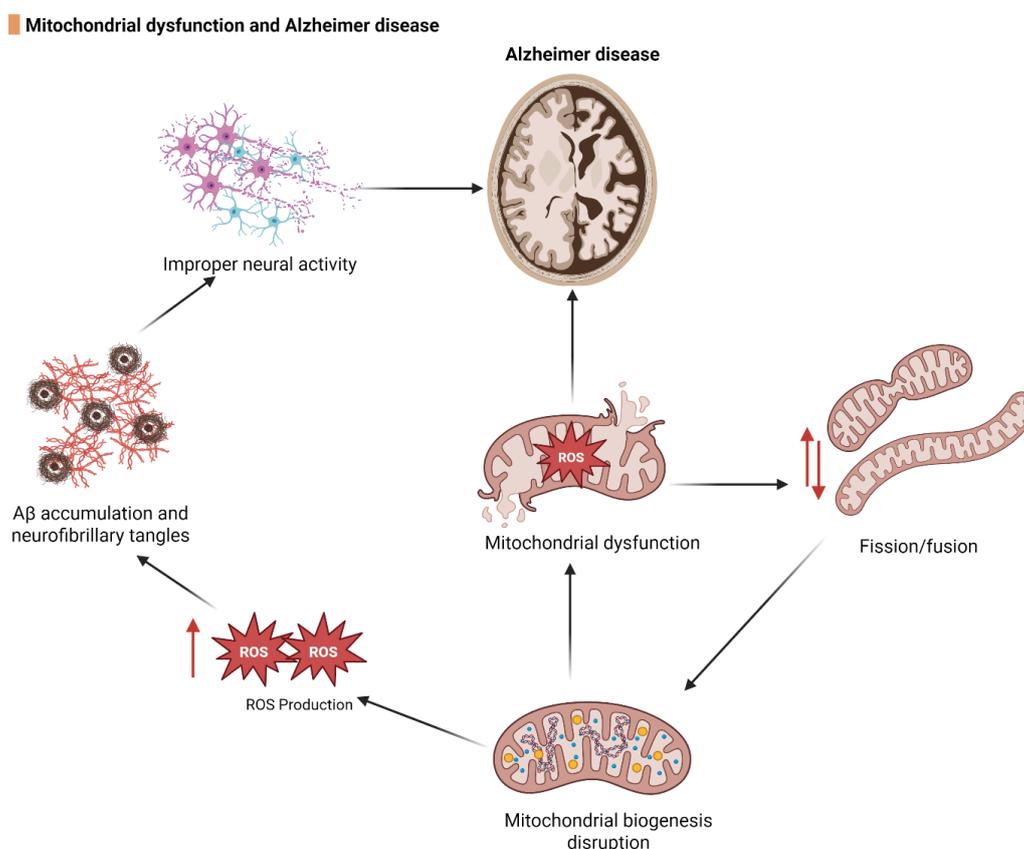


Figure 2. How Dysregulated Mitochondrial Dynamics Contribute to Alzheimer's Disease Pathology.

Note. Under normal conditions, there are no signs of abnormality; mitochondrial fusion and fission balance each other to maintain mitochondrial integrity and proper distribution, ensuring normal energy production. However, this balance shifts toward excessive mitochondrial fission in abnormal conditions. Increased mitochondrial fragmentation leads to abnormal intracellular distribution resulting from these defects, which disturb neuronal energy homeostasis, changes, and prompt neuronal apoptosis, thereby accelerating cognitive decline and showing primary signs of Alzheimer's disease.

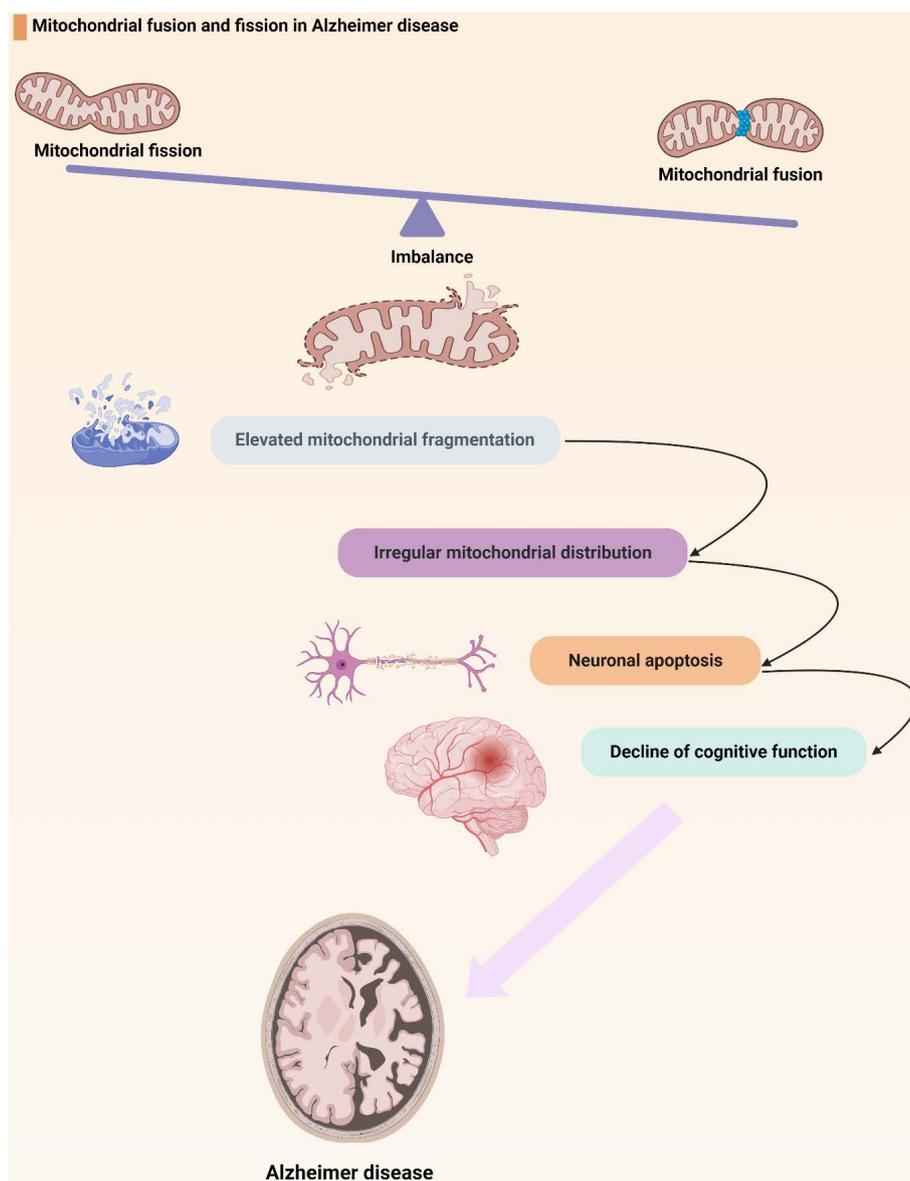


Figure 3. Physiological Associations Between Mitochondrial Dysfunction and the Initiation of Alzheimer's Disease

Note. A β : Beta-amyloid; ROS: Reactive oxygen species. Mitochondrial fission and fusion, along with affected mitochondrial biogenesis, lead to increased ROS production and mitochondrial damage. That an abnormal state, as well as mitochondrial biogenesis that does not work right, cause the mitochondria to make more ROS and get hurt. More ROS leads to more A β accumulation and the formation of neurofibrillary tangles, causing neurons to function less effectively and leading to abnormal behavior. The cycle of oxidative stress, mitochondrial damage, and protein aggregation causes severe neurodegeneration, which is a sign of Alzheimer's disease.

active transition metal ions, and decreased antioxidant levels.^{21,22} According to research, ROS can damage cells in diseases that impair the brain's ability to function.²³ In AD, ROS causes the accumulation of Ab proteins, thereby disrupting lysosomal membrane integrity and killing neurons.²⁴ In AD, cytochrome c oxidase deficiency is the most common problem with the mitochondrial electron transport chain, leading to more ROS production, less energy storage, and problems with energy metabolism.²⁵ Additionally, ROS inhibits phosphatase 2A,²⁶ thereby facilitating the activation of glycogen synthase kinase 3 β (GSK-3 β), a kinase involved in the phosphorylation of tau. Increased GSK-3 β activation may result in tau hyperphosphorylation and neurofibrillary lesions in AD.²⁷ Oxidized biomolecules from ROS (e.g., the phosphorylation of tau) are highly stable and are often

used as ROS indicators. In addition, ROS can be indirectly measured by looking at the levels of antioxidants or the activity of enzymes. In AD, there is often an imbalance in the OS and an increase in by-products. Some studies indicate that AD markedly elevates lipid peroxidation, a process in which ROS target lipids, generating products via a free radical chain reaction.^{28,29} It is also essential to monitor malondialdehyde, a key marker of OS and lipid peroxidation, at regular intervals. It is a straightforward and cost-effective method for tracking how AD is worsening and how well treatment is working. Protein carbonyls, 3-nitrotyrosine, TBARS, free fatty acid release, isoprostane and neuroprostane formation, 2-propen-1-al (acrolein), 4-hydroxy-2-trans-nonenal, advanced glycation end products, and 8-OH-2'-deoxyguanosine (8-OH-guanos) are a few common OS markers that

are found in biological samples. It should be noted that elevated levels of toxic carbonyls, 3-nitrotyrosine, and 4-hydroxy-2-trans-nonenal are initial alterations in AD following OS.^{30,31} Based on reports, oxidative and nitrosative stress can alter key biological molecules, including proteins, lipids, and nucleic acids. When oxygen is only partially reduced, it generates ROS, including the superoxide radical anion (O_2^-), hydrogen peroxide (H_2O_2), the hydroxyl radical ($HO\bullet$), nitric oxide (NO), and peroxynitrite (ONOO⁻). Oxidative phosphorylation in mitochondria is a primary source of free radicals, as electron leakage from the mitochondrial electron transport chain results in O_2 production.³² Moreover, progressive mitochondrial dysfunction is a principal catalyst of ROS generation in aging and AD, as well as a significant target of oxidative damage.³³ Some studies have linked mitochondrial dysfunction to altered ROS processing in AD.^{34,35} In addition, adding Ab oligomers to the bilayer can generate ROS, which can then damage membrane lipids, proteins, and nucleic acids inside the cell.^{36,37} OS is also associated with mitochondrial function, as ROS can impair mitochondrial function. Decreasing ROS through dietary modifications, physical activity, and antioxidant supplementation can safeguard mitochondrial function in the brain by mitigating oxidative stress and associated damage (Figure 3).

Current and Conventional Therapeutic Interventions

Current therapeutic interventions in AD encompass several approaches, reflecting ongoing trends, challenges, and emerging prospects. Despite significant advances in understanding the pathophysiology of the disease, a definitive cure remains unavailable.³⁸

Approved and Traditional Drugs

Cholinesterase inhibitors, including donepezil, rivastigmine, and galantamine, function by inhibiting the breakdown of acetylcholine, thereby enhancing cognitive performance in patients with mild-to-moderate AD.³⁹ Meanwhile, the N-methyl-D-aspartate receptor antagonist memantine modulates glutamate transmission, helping mitigate excitotoxicity in individuals with moderate to severe disease.⁴⁰ Although these pharmacological agents offer symptomatic management, they do not alter the underlying trajectory of disease progression. Recent advancements in formulation, such as Namzaric (a combination of memantine and donepezil) and transdermal donepezil patches, strive to promote patient adherence while minimizing the adverse effects.⁴¹

Targeted and Next-Generation Drugs

Recent research is increasingly focused on targeted interventions for amyloid and tau pathologies. Monoclonal Abs, including aducanumab, lecanemab, and donanemab, are designed to facilitate the clearance of A β plaques and thereby decelerate AD progression. However, the clinical efficacy of these therapies remains contentious

within the scientific community.⁴² Sodium oligomannate (GV-971) modulates the gut microbiome, thereby reducing neuroinflammation, a factor increasingly recognized as a key driver of neurodegenerative disease pathology.⁴³ In addition, selective inhibitors of molecular pathways (e.g., GSK-3 β , DYRK1A, and HDAC6) and dual-target compounds (e.g., ladostigil) are being used to enhance specificity while decreasing toxicity.⁴⁴ Moreover, innovative therapeutic strategies are emerging, such as proteolysis-targeting chimeras and protein-protein interaction inhibitors. These strategies are under investigation for their potential to selectively degrade pathogenic proteins associated with neurodegeneration. It is noteworthy that these advancements signal a promising horizon for addressing the underlying mechanisms of AD and related disorders.⁴⁵

Combination and Multifactorial Therapies

Given the multifactorial etiology of AD, there is a trend toward employing combination therapies to enhance treatment efficacy. Notable pairings (e.g., Varoglutamstat with Aducanumab—which specifically target neurotoxic N3pE peptides) are yielding promising preliminary results.⁴⁶ Additionally, the integration of radiotherapy, histone deacetylase inhibitors, and nutritional supplements, along with neuroprotective agents, appears to confer a synergistic advantage. These multitarget approaches not only aim to boost therapeutic outcomes but also may mitigate adverse effects and help prevent the development of drug resistance.⁴⁷

Complementary and Alternative Medicine

Both Mediterranean and ketogenic dietary patterns have been shown to inhibit amyloidosis and tau phosphorylation processes, which are pivotal in neurodegenerative diseases.⁴⁸ Acupuncture has been demonstrated to modulate synaptic protein expression and enhance neural plasticity, potentially offering therapeutic benefits for cognitive function.⁴⁹ Further, some practices, such as Kirtan Kriya yoga, have been linked to reductions in OS, subsequently improving both mental performance and mood regulation. Furthermore, herbal extracts from Ginkgo biloba, Bacopa monnieri, and Curcuma longa, as well as other plants rich in polyphenolic compounds, exhibit essential anti-amyloidogenic and anti-inflammatory properties.⁵⁰ Finally, routine physical exercise is associated with increased cerebral blood flow and neuroprotection, contributing to overall cognitive resilience.⁵¹

Stem Cell Therapy

It represents an innovative and promising intervention in the regenerative medicine landscape, particularly in addressing neurodegenerative disorders, such as AD. Various stem cell types, including pluripotent, mesenchymal, and neural stem cells, have demonstrated significant potential for repairing and replacing damaged

neuronal tissue. Key characteristics of stem cells drive their therapeutic application, notably their capacity for self-renewal and differentiation into diverse specialized cell types. They are classified into totipotent, pluripotent, multipotent, and unipotent types.⁵² In neurodegeneration, this type of therapy aims to utilize stem cells and their derivatives to restore neuronal function. Mesenchymal stem cells (MSCs), which are multipotent and can be sourced from various tissues (e.g., bone marrow, adipose tissue, umbilical cord, placenta, amniotic fluid, and dental pulp) have garnered particular interest.⁵³ These cells secrete a range of anti-inflammatory and neurotrophic factors, as well as exosomes (EXOs) that facilitate nervous system repair. In preclinical AD models, MSC transplantation has been shown to reduce amyloid plaques while improving cognitive function. Clinical trials have begun to reveal modest cognitive stabilization and improvements in patients treated with MSCs. The evidence suggests efficacy in other neurodegenerative conditions, such as Parkinson's disease, amyotrophic lateral sclerosis, and multiple sclerosis, through different mechanisms, including neuroprotection, reduced inflammation, remyelination, and enhanced neural function.⁵⁴ Likewise, some studies have explored the genetic modification of MSCs to promote differentiation into cholinergic neurons, with subsequent transplantation into cortical regions leading to synaptic reconstruction and improvements in memory processes. Establishing standardized protocols for the extraction, cultivation, transplantation, and long-term safety assessment of MSCs is crucial to extending these therapeutic strategies.⁵⁵ Nevertheless, several challenges must be addressed before implementing stem cell therapies, notably concerns about safety, tumorigenesis, and effective penetration into brain tissue.⁵⁶ Despite advancements, hurdles persist in achieving targeted differentiation and functional integration within host neural networks and ensuring the long-term safety of the interventions. Progress in imaging techniques, cellular engineering, and clinical trial methodologies may ultimately facilitate the development of safe and effective stem cell therapies for AD and other neurodegenerative conditions.⁵⁷

Extracellular Vesicles as an Innovative and Cell-Free Therapy

Research indicates that intercellular communication predominantly occurs through paracrine signaling, in which cells release EVs into the extracellular matrix. These EVs play a critical role in mediating signaling cascades and modulating cellular responses within the local microenvironment.⁵⁸ Molecular and ultrastructural analyses categorize these vesicles into three primary subtypes: apoptotic bodies, EXOs, and macrovesicles. Apoptotic bodies range from 1,000 nm to 5,000 nm in diameter. In addition, macrovesicles are approximately 100 nm to 1,000 nm, and EXOs are smaller, measuring between 50 nm and 200 nm.⁵⁹ Among them, EXOs are

of particular interest in the context of EV research due to their significant role in advancing cell-based therapies and drug-delivery systems. These nanoscale EVs exhibit intricate genomic, proteomic, and lipidomic profiles and can be isolated from various biofluids, thereby enhancing their utility as targeted delivery vesicles in regenerative medicine applications.⁶⁰⁻⁶² Moreover, their intrinsic biocompatibility and capacity to facilitate intercellular communication render them exceptionally valuable in therapeutic settings.^{63,64} Importantly, nearly all cell types can synthesize and secrete EVs in both physiological and pathological contexts.^{65,66} Further, EVs have nanoscale dimensions and a lipid bilayer, enabling them to cross the blood-brain barrier (BBB) and deliver biomolecules (e.g., proteins and nucleic acids) directly to neuronal cells.⁶⁷ This makes EVs efficient drug-delivery systems in AD, reducing side effects and targeting damaged areas. Although altered EV function can contribute to AD progression, modifying their cargo can transform them into therapeutic carriers that reduce inflammation and protein accumulation, thereby enhancing cognitive functions. Likewise, EVs loaded with micro ribonucleic acids (miRNAs) and short interfering RNAs targeting genes (e.g., *BACE1* and *tau*) demonstrate promise for treating AD by silencing harmful genes.⁶⁸ Preclinical studies have successfully engineered EVs for targeted brain delivery of small RNAs, mitigating pathological protein aggregation. Additionally, EVs derived from mesenchymal and neuronal stem cells provide neurotrophic and anti-inflammatory benefits, making this cell-free strategy safer than traditional cell transplants. Furthermore, these vesicles serve as biomarkers for early AD diagnosis and disease monitoring, encapsulating pathogenic molecules detectable in biofluids.⁶⁹ Research highlights the role of MSC-derived EVs, particularly those overexpressing miR-146a, in reducing inflammation while improving cognitive function in murine models.⁷⁰ As AD progresses, amyloid-beta ($A\beta$) and tau aggregates accumulate in predictable patterns, with EVs mediating intercellular communication and the transmission of disease factors. Inhibiting EV release has shown potential to alleviate AD symptoms, emphasizing their key role in the development of the disease.⁶⁷ Similarly, emerging data suggest that MSC-derived EVs can clear pathological proteins (e.g., $A\beta$ plaques and hyperphosphorylated tau), activate autophagic pathways, and modulate proteins and microRNAs (miRNAs) essential for neuronal viability. In addition, these vesicles enhance autophagy, regulate neuroprotective proteins, such as sphingosine 1-phosphate (S1P) and neprilysin (NEP), and promote anti-inflammatory responses. S1P signaling enhances cognitive function and reduces $A\beta$ deposition, while NEP facilitates amyloid degradation. Additionally, EVs transfer miRNAs that mitigate neuroinflammation and support neuronal health. They also alleviate OS by delivering antioxidative enzymes and activating the nuclear factor erythroid 2-related factor 2 pathway, promoting a

reparative microglial state. Further, mesenchymal EVs prevent neuronal apoptosis, promote neurogenesis, and enhance synaptic function, thereby improving cognitive outcomes. Overall, MSC-derived EVs represent a promising therapeutic approach for AD, as they reduce neuroinflammation and protect neurons.⁷¹

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Neuroinflammation

Neuroinflammation plays a crucial role in AD development and progression, mainly due to the dysregulated activation of microglia. In a healthy brain, microglia remain quiescent, but in AD, they are activated by damage-associated and pathogen-associated molecular patterns via pattern recognition receptors. The nucleotide oligomerization domain-like receptor protein 3 (NLRP3) inflammasome is particularly important, as its activation in microglia leads to increased interleukin (IL)-1 β levels. Inhibiting NLRP3 can reduce A β accumulation, lower levels of inflammatory cytokines, and improve cognitive deficits while also decreasing tau hyperphosphorylation

(Figure 4).^{72,73} Research has shown that MSC-EVs enhance the expression of genes involved in synaptic plasticity and memory, thereby contributing to neuroprotection. Furthermore, they improve calcium homeostasis and neuronal function while reducing neuronal apoptosis through the phosphatase and tensin homologue-phosphatidylinositol 3-kinase/protein kinase B pathway.⁷⁴ The anti-inflammatory cytokine IL-10 in MSC-EVs helps shift macrophage polarization from a pro-inflammatory to an anti-inflammatory phenotype, thereby improving immune responses in the central nervous system (CNS). Moreover, MSC-EVs alleviate OS by reducing inducible NO synthase expression and delivering catalase, thereby enhancing neuronal survival. Additionally, the activation of the nuclear factor erythroid 2-related factor 2 signaling pathway by MSC-EVs is vital for maintaining redox balance and reducing inflammation, further supporting neuronal function. Likewise, MSC-EVs effectively target neuroinflammation, modulate immune responses and microglial function, and attenuate OS. These mechanisms collectively facilitate neural regeneration, underscoring

Neuro Inflammation mechanism in Alzheimer Disease

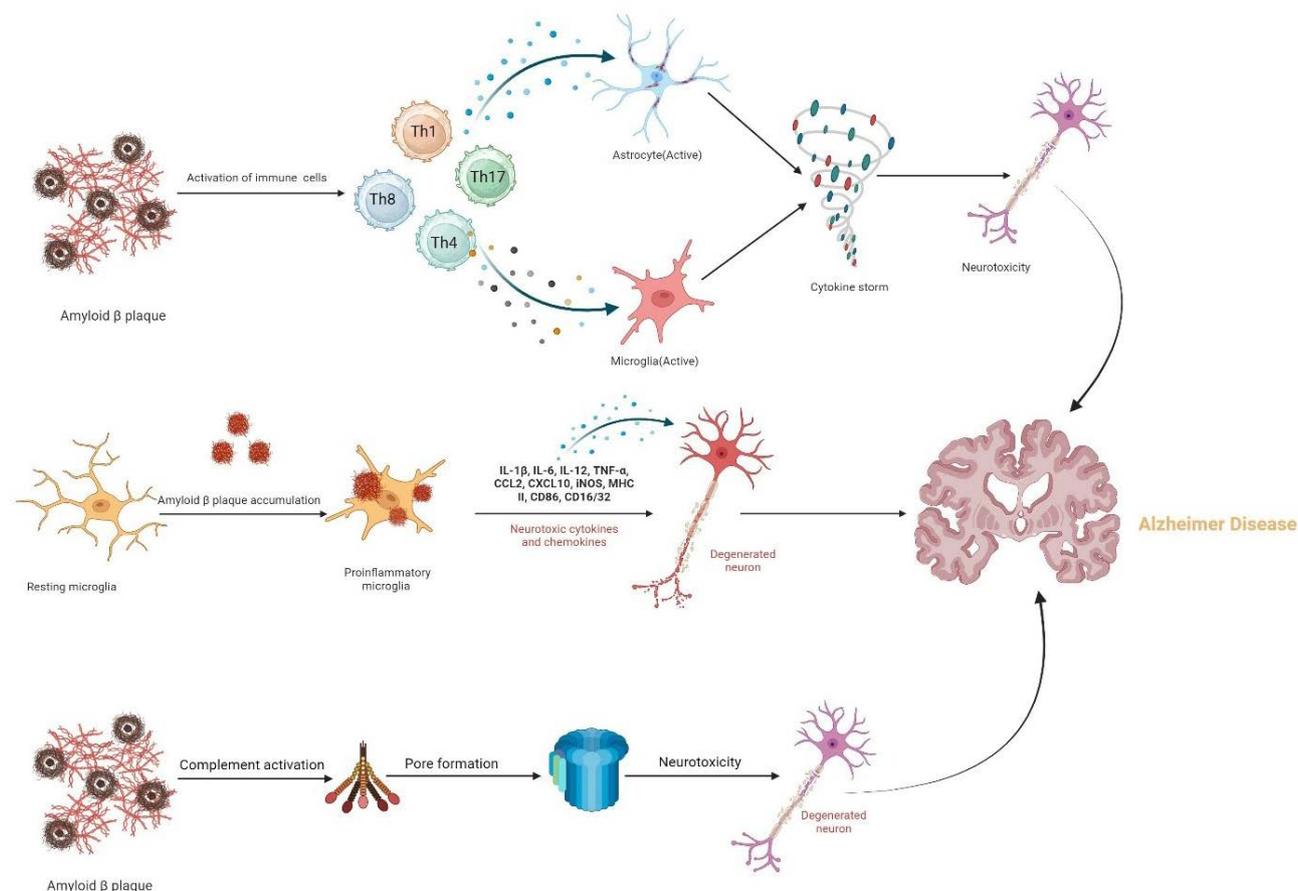


Figure 4. Different Ways of Showing That Neuroinflammation Happens in Alzheimer's Disease

Note. Th: T helper; IL: Interleukin; TNF- α : Tumor necrosis factor-alpha; A β : Beta-amyloid. The diagram illustrates three primary mechanisms by which the accumulation of A β plaques leads to neurotoxicity and ultimately contributes to the development of Alzheimer's disease: Activation of immune cells. A β plaques stimulate various types of Th cells (including Th1, Th4, Th8, and Th17) to become active, thereby triggering astrocytes and microglia, leading to a cytokine storm and neurotoxicity. Microglial transition: Resting microglia collect A β plaques and transition into proinflammatory microglia that release neurotoxic cytokines and chemokines (e.g., IL-1 β , IL-6, IL-12, TNF- α , CCL12, and the like), causing the neuron to degenerate. Activation of the complement system: A β plaques cause the complement system to become active, leading to the formation of pores and neurotoxicity and neuronal death.

the potential of MSC-EVs to advance AD therapies.⁷¹

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Immunological Responses

MSCs are particularly essential for their potent immunomodulatory properties,^{74,75} making them promising therapeutic candidates for immune homeostasis and regenerative medicine.^{75,76} They efficiently modulate innate and adaptive immunity^{77,78} via the secretion of bioactive molecules and direct cell-to-cell interactions with immune cells.^{77,78} Similarly, MSCs and EVs derived from them are highly suitable for treating certain inflammatory and autoimmune disorders due to their distinctive characteristics, which enable them to reduce inflammation, regenerate tissue, and maintain immunological homeostasis.^{79,80} As an alternative to MSCs, MSC-EV offers numerous benefits, including reduced immunogenicity and an enhanced safety profile,^{82,83} as well as the capacity to cross biological barriers.⁸¹ Additionally, the MSC-EV application circumvents immunological rejection, entrapment of the lung microvasculature, and ectopic tumor formation associated with stem cells.^{82,83} The creation of EV as therapeutic regimens is amply justified, if not called for, by these benefits and mounting evidence for the therapeutic effects of MSC-EV.⁸³ According to recent research, treatment with MSCs applied in neurodegenerative illnesses such as AD is safe and efficient, even when more invasive delivery methods are utilized, such as intracerebral injection.⁸⁴ It is challenging to make apples-to-apples comparisons of clinical trial results, as serious heterogeneity remains in the sources, doses, and modes of administration of MSCs. Autologous bone marrow-derived MSCs and adipose tissue-derived MSCs are the most frequently examined cells for neurological therapy. However, over recent years, more attention has been paid to the umbilical cord as a promising source of MSCs.⁸⁴ Umbilical cord-derived MSCs possess similar characteristics (e.g., low immunogenicity) and certain benefits over other sources; the primary benefit is that they can be obtained in bulk quantities from a disposable source (the post-birth umbilical cord) without discomfort or harm to mother or child.⁸⁵ It should be noted that MSCs come from diverse sources, can be easily isolated and amplified, have low immunogenicity, have low teratoma formation potential, have high migratory and interactive potential, modulate diverse biological processes, and are, therefore, a promising option for autologous and allogeneic cell therapies in neurological disease based on preclinical studies.⁸⁶ Nonetheless, the QC and reproducibility of MSC therapy in clinical trials are more contentious.

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Inflammatory Cytokines and Chemokines

MSC-derived EVs express diverse adhesion molecules (CD29, CD44, and CD73), enabling them to target inflamed and damaged tissues. In the mouse intracerebral

hemorrhage model, MSC-EVs were accumulated in the damaged brains.⁸⁷ Likewise, EVs tagged with the RVG protein targeted neuronal cells more specifically and showed better therapeutic effects, with greater attenuation of AD, compared to naïve EVs.⁸⁸ Kooijman et al used glycosylphosphatidylinositol. This glycolipid is incorporated into the EV's membrane during biogenesis to deliver chemokine receptors, enzymes, Abs, and signaling molecules, thereby enhancing tropism and therapeutic effect.⁸⁹ MSC-EV membranes contain high levels of cholesterol, sphingomyelin, ceramide, and lipid raft proteins.⁹⁰ that enables membrane fusion with target cells and tracking of MSC-EVs through the body, regardless of biological barriers.⁹¹ It was suggested that MSC-EVs, like tumor-derived EVs, cross the BBB and that transcytosis is the primary underlying mechanism.⁹² The endothelial recycling endocytic pathway is involved in the transcellular transport of EVs.⁹³ After MSC-EVs have arrived at their target cells, they can communicate with each other via receptor-ligand interactions or be endocytosed to deliver their contents.⁹⁴ All MSC-EVs contain MSC-derived bioactive molecules (messenger RNA [mRNA], miRNAs, enzymes, cytokines, chemokines, immunomodulatory factors, and growth factors) that regulate immune cell phenotype, function, survival, and homing.⁹⁵ The immunosuppressive action of MSC-EVs was comparable to that of MSC transplantation.⁹¹ It is noteworthy that MSC-derived EVs, a cell-free product, circumvent safety concerns associated with the long-term survival of MSCs, including uncontrollable differentiation, malignancy, and allogeneic immune response-induced rejection in MHC-mismatched patients.⁹⁵ The *in vitro* preconditioning of MSCs can modulate the content of MSC-EVs, thereby producing a disease-specific immunosuppressive product for cell-free therapy of inflammatory and autoimmune diseases.⁹⁵ MSCs contain catalase, an active enzyme that helps protect cells against OS. Catalase is crucial for MSC-EV anti-ROS-induced damage, as inactive catalase is incapable of suppressing ROS production in hippocampus neurons.⁹⁶ MSCs can also switch macrophages from M1 to M2, which are the primary sources of immunomodulatory cytokines, including arginase-1, A-degrading enzymes (NEP and insulin-degrading enzyme), transforming growth factor beta (TGF- β), IL-10, and TGF- α . In addition, MSC-EVs play a crucial role in treating AD by targeting A β pathology through distinct mechanisms. Adipose-derived MSC-EVs reduce A β 42 and A β 40 in the neural stem cells of AD mice, as well as both secreted and intracellular A β in human APP-expressing cells, potentially via NEP delivery.^{97,98} Bone-marrow MSC-EVs reduced A β plaques and dystrophic neurites in APP/PS1 mice, a model of AD, through NEP.⁹⁹ Additionally, the miRNA contents of MSC-EVs influence inflammation, synaptic density, and cognitive function. In animal models of AD, miR-146a reduces proinflammatory microglial activation by inhibiting nuclear factor kappa B in astrocytes,¹⁰⁰ whereas

miR-21-5p improves synaptic function while decreasing A β plaques.^{101,102}

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Microglial Activation

A neurodegenerative disease causes progressive neuron degeneration and death, leading to cognitive and behavioral dysfunction.¹⁰³ Glial cells, or neuroglia, are non-neuronal CNS cells. These cells can be divided into four types: astrocytes, microglia, oligodendrocytes, and NG2a-glia. They regulate brain plasticity, protect neurons, and maintain homeostasis.¹⁰⁴ Microglia, the immune cells of the CNS, maintain tissue homeostasis under physiological conditions.¹⁰⁵ Furthermore, activated microglia release pro-inflammatory cytokines after neuronal damage. These cytokines can result in neurotoxicity or induce neutrophil migration into the inflamed tissue.¹⁰⁵ Additionally, hyperactivated microglia harm healthy neural tissue and cause dying neurons to release alarmins and DAMPs, creating a “positive inflammatory loop” in the CNS that leads to ongoing neuron death.¹⁰⁵ Recent research by Ding et al indicates that the therapeutic effects of MSC-EVs on AD primarily result from the modulation of microglial function.¹⁰⁶ The cerebral accumulation of excessive A β is a prevalent pathogenic characteristic of AD, leading to cognitive impairment.¹⁰⁷ The intravenous infusion of EXOs from human umbilical cord-derived MSCs could diminish A β deposition and enhance spatial learning and memory performance in APP/PS1 transgenic mice, a model of AD.⁶² In their recent study, Bodart-Santos et al discovered that MSC-EVs mitigated neuronal damage in AD by alleviating OS-induced injury in hippocampal neurons.⁹⁶ Interactions among neurons, microglia, astrocytes, and oligodendrocytes can injure the CNS, leading to neuronal death and myelin damage.¹⁰⁸ Inflammation-activated astrocytes are being increasingly investigated for their role in contributing to neurodegenerative diseases.^{109,110} Several studies also reported that type 1 astrocytes (neurotoxic phenotype) and proinflammatory cytokines had a role in neurodegenerative diseases.^{111,112} Impaired astrocytes and activated microglia cooperated to reduce neuron survival in AD animal models.¹¹³ Impaired microglia have been shown to significantly contribute to neurodegeneration. The BBB prevents peripheral immune cells from entering the brain, making it an “immunologically privileged” organ. Microglial activation triggers neuroinflammation by releasing ROS, NO, and the inflammatory cytokines IL-1 β , IL-6, and tumor necrosis factor (TNF).¹¹⁴ Likewise, activated microglia, proinflammatory mediators, and increased OS, and nitrosative stress are associated with chronic inflammation and neurodegenerative diseases.¹¹⁵ Specific signaling molecules, including nucleotides, cytokines, and chemokines, are released by neurons undergoing apoptosis.¹¹⁶ Astrocytes also control neuroinflammatory events.¹¹⁷ Inflammatory stimuli appear to be mostly

passive for microglia. In addition, microglial activation drives the transformation of astrocytes into toxic A1 cells. A1 astrocytes are abundant in neurodegenerative diseases, especially AD.¹¹⁸ Although overactivated macrophages and microglia can cause immunotoxicity, they play a crucial role in the process. Microglia and macrophages can remove myelin debris from lesions, thereby inhibiting the development of oligodendrocyte precursor cells.¹¹⁹ MSC-EXO skewed microglia to the M2 immunosuppressive phenotype. In A/PP/PS1 mice treated with MSC-Exos, M2 microglial cells expressing chitinase 3-like 3, arginase-1, and MRC1 were increased in the brains.¹⁰⁶ In M2 cells, A-degrading enzymes (NEP and IDE) and anti-inflammatory cytokines (IL-10 and TGF- β) could reduce A deposition and inflammation [61]. Moreover, MSC-Exo treatment of APP/PS1 mice resulted in increased NEP, IDE, IL-10, and TGF- α and decreased inflammatory cytokines (TNF- α and IL-1 β) in the brain, reflecting a shift from inflammatory M1 to anti-inflammatory M2 microglia.¹⁰⁶ Myelin basic protein is a myelin protein present in macrophages and microglia.¹²⁰ Damaged axons do not provide the requisite structure and physiology for remyelination by CNS precursor cells.¹²¹ Neurodegenerative disorders (e.g., AD) have shown MSC-EVs to have powerful therapeutic potential for treating various diseases by delivering therapeutic cargo, decreasing inflammation, enhancing tissue repair, and improving neuronal survival and function.¹²²

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Mitochondrial Condition

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Mitochondrial Fusion and Fission

Mitochondrial dysfunction is a hallmark of AD. However, current mitophagy inducers are toxic and do not effectively accumulate in the brain.¹²³ Numerous studies have shown that this type of dysfunction plays a crucial role in AD initiation and progression, occurring prior to the formation of amyloid plaques and NFTs.¹²⁴ Zhang et al investigated protective effects using human umbilical cord-derived MC-EVs in an in vitro model of AD and analyzed the associated mitochondrial mechanisms. Xu et al also generated nanosized MSC-derived EVs (MSC-EV-SHP2) that expressed high amounts of tyrosine phosphatase-2 (SHP2). Due to the excellent BBB penetration potential, MSC-EV-SHP2 could effectively deliver SHP2 to AD mice. Furthermore, it significantly reduced NLRP3-activated inflammasomes and apoptosis associated with mitochondrial damage in neural cells. Mitophagy further decreased inflammation and neuronal death in AD mouse models. Accordingly, it is an efficient EV-engineering strategy to promote mitophagy as a therapy in AD patients.¹²⁵ Dynamin-related protein 1 (Drp1) is one essential protein that is involved in mitochondrial fission. It is found in all living things, from yeast to mammals. It should be noted that this protein is vital for mitochondrial fission.^{126,127} Loson et al reported that

resident protein receptors (e.g., Mff, Fis1, and MiD49/51) recruit Drp1 to the outer mitochondrial membrane in order to facilitate mitochondrial fission. Drp1 is essential for mitochondrial division, size, shape, and distribution in neurons, from the cell body to axons, dendrites, and nerve terminals. Furthermore, the Drp1 protein has a significant effect on mitochondrial movement, fusion, and the engulfment of other mitochondria. More precisely, it is a mitochondrial fission factor that breaks up mitochondria and makes fusion easier when it is turned down.¹²⁸ Drp1 imbalance can lead to mitochondrial dysfunction, which is a problem in AD and other neurodegenerative diseases.^{129,130} Moreover, diseases that cause excessive mitochondrial fragmentation can raise Drp1 levels, leading to mitochondrial and nerve damage. In vitro AD models demonstrate that mitochondrial fission persists longer and has detrimental effects. When overexpressed or treated with Abs, APP can cause mitochondria to break apart and alter their distribution, resulting in problems with synapses in neuronal cultures.^{129,131} Fission and fusion processes are linked to mitochondrial metabolism.¹⁶ Mitochondrial fission is increased, thereby reducing mitochondrial oxidative phosphorylation (OXPHOS), and cell homeostasis requires a balance between fission and fusion.^{132,133} Although mitochondrial fusion is not required for cell viability, it is necessary for normal growth. MSCs can rescue OXPHOS and alter morphology by promoting metabolic activities and ROS production.¹⁶ Mitochondrial fusion genes (*mfn1* and *mfn2*) and optic atrophy 1 can be induced to protect cells from extracellular damage by enhancing respiration characteristics.¹³⁴ Additionally, MSCs can suppress mitochondrial apoptosis and oxidative damage in damaged cells by inhibiting the release of cytochrome into the cytoplasm. The cells secrete cytokines and growth factors that enhance anti-apoptotic proteins (BCL-XL and BCL-2) while decreasing pro-apoptotic proteins (BAX, BAK, and BAD) and cytochrome, thereby minimizing mitochondrial injury.¹³⁵

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Mitochondrial Quality Control

Mitochondrial quality management necessitates monitoring and preventive measures at multiple levels in order to ensure safety and optimal function. QC happens at the molecular, organelle, and cellular levels. At the molecular level, a proteolytic mechanism in the mitochondrial matrix and inner membrane breaks down damaged and misfolded proteins and disassembles protein aggregates for proteolysis.¹³⁶ In addition, the ubiquitin-proteasome system in the cytosol controls the quality of mitochondrial proteins.¹³⁷ Further, mitochondrial fusion and fission provide additional protection against mitochondrial damage at the organelle level. The combination of dysfunctional mitochondria and healthy mitochondria results in a mixture of healthy and defective mitochondria.^{138,139} Severely damaged mitochondria are

occasionally split apart by fission, leading to their removal from the autophagy-lysosomal system via mitophagy.¹⁴⁰ If the QC processes at the molecular and organelle levels are not up to par, or if the damage is too severe for them to handle, damaged mitochondria can rupture and release pro-apoptotic proteins, causing apoptosis and cell death. To maintain the health and proper functioning of mitochondria at the molecular, organelle, and cellular levels, it is essential to perform QC on them. QC that is not good enough at every step causes mitochondria to malfunction and damaged mitochondria to accumulate in unexpected ways. An imbalance in mitochondrial dynamics, diminished axonal transport, and insufficient mitochondrial phagocytosis have been identified in AD neurons, potentially hindering the effective elimination of dysfunctional mitochondria. Thus, bad QC could lead to mitochondrial disease in AD.¹⁴¹ In AD, mitochondrial QC is impaired at the molecular level. The mitochondrial sequence protease presequence protease (PreP) degrades Ab within mitochondria, thereby diminishing its toxic impact on mitochondrial function.^{142,143} Studies on the brains of AD patients and mice support the idea that enhanced ROS production impairs PreP proteolytic activity, leading to Ab accumulation and mitochondrial toxicity. Additionally, a decline in proteasome activity would affect the QC of other peptides and mitochondrial proteins.^{144,145} It should be noted that the mitochondrial dysfunction detected in AD can be exacerbated by the toxic impact of the damaged peptides and mitochondrial proteins that accumulate accordingly.¹⁴⁶

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Mitochondria and Beta-Amyloid

AD brains demonstrate problems with their mitochondria, including reduced function, changes in dynamics and transport, increased mitochondrial DNA mutations, altered enzyme activity, and abnormal gene expression.¹⁴⁷ Reports indicate that the accumulation of APP and A β peptides is a significant contributor to mitochondrial toxicity. Purified mitochondria from patient brains and AD mouse models contain APP and A β .^{148,149} According to research, A β interacts with mitochondrial matrix proteins ABAD and cyclophilin D, which have harmful effects.^{150,151} Research further represents that the abnormal accumulation of A β in synaptic mitochondria may lead to early synaptic dysfunction in AD.¹⁵² Moreover, mitochondrial dysfunction and Ab accumulation in the brains of people with AD are linked to cognitive decline.¹⁵³ Mitochondria are accessed via the mitochondria-associated membranes⁸⁷ or the translocase of the outer membrane complex. Hansson Petersen et al found that mitochondria internalize and absorb extracellular A β .¹⁵⁴ The findings of a recent study revealed that only the proximity of A β plaques in a living AD mouse model reduced mitochondrial membrane potential and led to the emergence of dystrophic and fragmented mitochondria, suggesting that A β plaques may serve as the focal sources

of mitochondrial A β accumulation and toxicity.¹⁵⁵ These results suggest that APP and A β accumulation in the mitochondrial compartment may cause mitochondrial dysfunction by changing mitochondrial dynamics and transport. Hence, removing pathological proteins (e.g., A β plaques and hyperphosphorylated tau), which induce synaptic disruption and neuronal degeneration, is crucial for treating AD.¹⁵⁶ Research also indicates that natural MSC-EVs may help clear pathogenic proteins by modulating autophagy and regulating key protein levels in AD development, including those of hereditary proteins and miRNAs. miR, a 22-24 nucleotide non-coding RNA present in EVs, silences mRNA transcripts by attaching to the 3' untranslated region.¹⁵⁷ Decreased v-ATPase activity and selective accumulation of A β /A β precursor protein (APP) in larger de-acidified autolysosomes.¹⁵⁸ Additionally, autophagy activation effectively scavenges A β plaques, suggesting that it is a promising method for preventing AD by removing aberrant protein buildup.¹⁵⁸ MSCs and MSC-EVs can activate autophagy, thereby reducing aberrant protein levels while improving AD.^{159,160} In addition to activating autophagy, MSC-EVs can regulate the production of a critical protein that can eliminate aberrant proteins. Some researchers concluded that MSC-EVs can treat AD by regulating S1P expression, which promotes autophagy, protects neurons, modulates glial cell inflammatory responses, and inhibits apoptosis, thereby preventing the pathological changes associated with AD.¹⁶¹ Cui et al used EXOs from hypoxia-preconditioned mesenchymal stromal cells to test cognitive rehabilitation in an APP/PS1 model of AD. MSCs and hypoxia-preconditioned MSC-derived EXOs reduced intracellular and extracellular A β oligomer deposits; however, MSCs were more effective at reversing learning and memory impairments.^{101,162} This was achieved by minimizing pro-inflammatory cytokines (IL-1 β and TNF- α) while maximizing anti-inflammatory cytokines (IL-4 and IL-10). Furthermore, EXOs minimized inflammation by inhibiting the activity of astrocytes and microglia. The obtained data confirmed that MSC-EVs enhance the memory of AD mice by activating the SphK/S1P signaling pathway and reducing A β deposition through the modulation of autophagy. In addition, NEP is a membrane-bound metallopeptidase that cleaves neuropeptides and amyloid proteins, making it a possible target for AD therapy.¹⁶³ Studies on living and nonliving cells have shown that NEP, present on MSC-EVs, can reduce A β accumulation.^{98,99} Research reported that MSC-EVs might be able to get rid of harmful proteins using their inherited miRs. BACE1 or BACE1 inhibitors could be used to treat AD because β -secretase cleavage starts the breakdown of APP.¹⁶⁴ According to recent studies, BACE1 controls AD progression through miRs.¹⁶⁵ Sha et al¹⁶⁶ found that BMSC-EVs deliver miR-29c-3p to neurons, thereby suppressing BACE1 expression and activating the Wnt/ β -catenin pathway, which has a positive effect on AD.¹⁶⁷ Likewise, Jeong et al observed

that human umbilical cord MSC groups had significantly fewer amyloid plaques and lower BACE1 levels, as well as considerably more neurogenesis, compared to A β -injection groups.¹⁶⁸

Mesenchymal Stem Cell-Derived Extracellular Vesicles and Mitochondria and Tau Protein

Tau is an essential protein that is linked to microtubules and has a significant impact on neuronal function. This protein has six isoforms in adult human brains, all derived from the same gene via alternative mRNA splicing.¹⁶⁹ A proline-rich region exists between the microtubule-binding domain and the projection domain, containing several phosphorylation sites.^{170,171} The proline-rich tau region interacts with the microtubule surface, thereby stabilizing it.¹⁷² As the principal posttranslational modification of tau, phosphorylation profoundly influences its dynamic equilibrium with microtubules.^{173,174} Moreover, the phosphorylation of tau directed at serine and threonine has a direct effect on how well it binds to microtubules.¹⁷⁴ When tau does not bind to microtubules properly, it clumps, forms fibrils, and stops working properly.¹⁷⁵ It should be noted that the microtubule network is vital for the movement of materials along axons. Microtubule dysfunction can lead to abnormal axonal transport and synaptic impairment. Tau alters the microtubule network, thereby affecting the movement of signaling molecules, trophic factors, and essential organelles (e.g., mitochondria) along axons. Additionally, Tau plays a role in critical cellular processes, including structural and regulatory functions.¹⁷⁶ Mitochondrial dysfunction is also essential in AD pathogenesis.¹⁷⁷ Reduced ATP production, elevated ROS levels, and dysfunctional OXPHOS complexes and antioxidant enzymes demonstrate that mitochondria are not functioning properly.¹⁷⁸ In cell culture and transgenic mouse studies, tau overexpression diminishes mitochondrial function by decreasing the activity of mitochondrial complexes and antioxidant enzymes while enhancing ATP synthesis and synaptic function.^{179,180} Furthermore, perinuclear mitochondrial distribution results in ATP depletion, OS, and synaptic dysfunction.^{34,181} The findings of a study revealed that hypophosphorylated tau directly interacts with Drp1, leading to mitochondrial fission and excessive fragmentation in postmortem brain tissues from AD patients and mice.¹⁸² Phosphorylated tau affects fission and interacts with voltage-dependent anion channel (VDAC) in AD brains, closing mitochondrial pores and preventing mitochondria from functioning properly.¹⁸² Research has shown that pathogenic Tau variants affect mitochondrial function by either interacting with VDAC or blocking Drp1-mediated fission. Tau is a microtubule-associated protein that keeps axonal microtubules stable. In addition, it controls the movement of membrane organelles along axons, such as mitochondria.¹⁸³ The overexpression of tau selectively blocks kinesin in neuroblastoma cell lines, primary cortical neurons, and retinal ganglion neurons.^{183,184} These

results indicate that tau and kinesin motors compete for binding to microtubules. Further, binding tau to microtubules can reverse the direction of dynein-motor movement, and the same process can cause kinesin to dissociate from microtubules, ultimately interrupting axonal transportation and neural connectivity.¹⁸⁵

AD is characterized by several key features, including hyperphosphorylation of the tau protein, which leads to the formation of NFTs, a buildup of A β , and mitochondrial dysfunction.¹⁸⁶ A key player in this intricate pathology is miR-132-3p, which is a miRNA that is highly expressed in brain tissue. Notably, diminished levels of miR-132-3p in the hippocampus correlate with impaired synaptogenesis and increased A β production, highlighting its vital role in neuronal health.¹⁸⁷ MSC-EVs emerge as powerful carriers of miRNAs, proteins, and metabolites that can effectively modulate tau and mitochondrial pathways in neurons affected by AD. Research reveals that miR-132-3p in MSC-EVs inhibits RAS P21 protein activator 1, thereby enhancing Ras signaling and activating the protein kinase B pathway. This cascade of events regulates GSK-3 β , which is crucial for controlling amyloid production and tau phosphorylation.¹⁶² Tau hyperphosphorylation is noticeably reduced by enriching MSC-EVs with miR-132-3p, thereby positively influencing mitochondrial dynamics and promoting neuronal survival against OS.¹⁸⁸ In the context of AD, mitochondrial dysfunction intensifies ROS production and disrupts axonal transport, leading to further neuronal degeneration. However, MSC-EVs can counteract these detrimental effects by releasing antioxidants and modulating the activity of GSK-3 β . Furthermore, MSC-EVs are strategically designed to target A β production by delivering short interfering RNAs against β -secretase and γ -secretase, alleviating mitochondrial stress, and preventing the progression of tau phosphorylation.¹⁸⁹ Most notably, MSC-EVs have the remarkable ability to cross the BBB while releasing anti-inflammatory factors, such as vascular endothelial growth factor and IL-10. This action not only supports mitochondrial function but also transforms the neuronal environment into a more anti-inflammatory state.¹⁹⁰ Therefore, MSC-EVs represent a groundbreaking, multi-target therapeutic strategy for AD, addressing its core pathological components and holding the potential to significantly alter the course of this devastating illness.¹⁶²

Conclusion and Forward-Looking Remarks

Given the complex and multifaceted nature of AD, it is crucial to develop innovative therapeutic strategies that effectively target its primary contributing factors and underlying mechanisms. MSC-EVs present a groundbreaking opportunity to modulate neuroinflammation, shifting the microglial response toward a protective anti-inflammatory state. Furthermore, these vesicles can help mitigate OS and restore mitochondrial function, including vital processes such as fusion, fission, and QC, thereby effectively addressing

the pathological hallmarks of AD. It is noteworthy that this cutting-edge, cell-free therapeutic platform not only promises to enhance immune response modulation but also holds the potential to reduce graft rejection and tackle tumorigenesis. While this approach is still in its initial stages, its development requires integrating advanced methodologies, robust foundational research, and rigorous preclinical trials in order to maximize its effectiveness and pave the way for clinical implementation. Overall, the future of AD treatment can lie within this innovative paradigm, opening new avenues for hope and healing in the fight against this devastating disease.

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Competing Interests

The authors declare no competing interests.

Ethical Approval

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References

- Zhang X, Liu H, Huang Y, Wang R. A meta-analysis of neurogenic exosomes in the diagnosis of Alzheimer's disease. *Heliyon*. 2023;9(10):e20604. doi: [10.1016/j.heliyon.2023.e20604](https://doi.org/10.1016/j.heliyon.2023.e20604)
- Zheng Q, Wang X. Alzheimer's disease: insights into pathology, molecular mechanisms, and therapy. *Protein Cell*. 2025;16(2):83-120. doi: [10.1093/procel/pwae026](https://doi.org/10.1093/procel/pwae026)
- Tarawneh R, Holtzman DM. The clinical problem of symptomatic Alzheimer disease and mild cognitive impairment. *Cold Spring Harb Perspect Med*. 2012;2(5):a006148. doi: [10.1101/cshperspect.a006148](https://doi.org/10.1101/cshperspect.a006148)
- Hill CV, Pike J. Commentary on Greenberg et al., "Prescribing anti-amyloid immunotherapies to treat Alzheimer's disease: fully informing patient decisions" [*Alzheimer's Dement*. 2023;9(4):e12426. doi.org/10.1002/trc2.12426]. *Alzheimers Dement (N Y)*. 2024;10(1):e12443. doi: [10.1002/trc2.12443](https://doi.org/10.1002/trc2.12443)
- Ryman DC, Acosta-Baena N, Aisen PS, Bird T, Danek A, Fox NC, et al. Symptom onset in autosomal dominant Alzheimer disease: a systematic review and meta-analysis. *Neurology*. 2014;83(3):253-60. doi: [10.1212/wnl.0000000000000596](https://doi.org/10.1212/wnl.0000000000000596)
- Dyrks T, Weidemann A, Multhaup G, Salbaum JM, Lemaire HG, Kang J, et al. Identification, transmembrane orientation and biogenesis of the amyloid A4 precursor of Alzheimer's disease. *EMBO J*. 1988;7(4):949-57. doi: [10.1002/j.1460-2075.1988.tb02900.x](https://doi.org/10.1002/j.1460-2075.1988.tb02900.x)
- Roher AE, Lowenson JD, Clarke S, Woods AS, Cotter RJ, Gowing E, et al. β -amyloid-(1-42) is a major component of cerebrovascular amyloid deposits: implications for the pathology of Alzheimer disease. *Proc Natl Acad Sci U S A*. 1993;90(22):10836-40. doi: [10.1073/pnas.90.22.10836](https://doi.org/10.1073/pnas.90.22.10836)
- Vershinin M, Carter BC, Razafsky DS, King SJ, Gross SP. Multiple-motor based transport and its regulation by tau.

- Proc Natl Acad Sci U S A. 2007;104(1):87-92. doi: [10.1073/pnas.0607919104](https://doi.org/10.1073/pnas.0607919104)
9. Wesseling H, Mair W, Kumar M, Schlaffner CN, Tang S, Beerepoot P, et al. Tau PTM profiles identify patient heterogeneity and stages of Alzheimer's disease. *Cell*. 2020;183(6):1699-713.e13. doi: [10.1016/j.cell.2020.10.029](https://doi.org/10.1016/j.cell.2020.10.029)
 10. Huggins LK, Min SH, Kaplan S, Wei J, Welsh-Bohmer K, Xu H. Meta-analysis of variations in association between APOE ε4 and Alzheimer's disease and related dementias across Hispanic regions of origin. *J Alzheimers Dis*. 2023;93(3):1095-109. doi: [10.3233/jad-221167](https://doi.org/10.3233/jad-221167)
 11. Yamazaki Y, Zhao N, Caulfield TR, Liu CC, Bu G. Apolipoprotein E and Alzheimer disease: pathobiology and targeting strategies. *Nat Rev Neurol*. 2019;15(9):501-18. doi: [10.1038/s41582-019-0228-7](https://doi.org/10.1038/s41582-019-0228-7)
 12. Liu CC, Zhao J, Fu Y, Inoue Y, Ren Y, Chen Y, et al. Peripheral apoE4 enhances Alzheimer's pathology and impairs cognition by compromising cerebrovascular function. *Nat Neurosci*. 2022;25(8):1020-33. doi: [10.1038/s41593-022-01127-0](https://doi.org/10.1038/s41593-022-01127-0)
 13. Mayeux R, Sano M. Treatment of Alzheimer's disease. *N Engl J Med*. 1999;341(22):1670-9. doi: [10.1056/nejm199911253412207](https://doi.org/10.1056/nejm199911253412207)
 14. Singh A, Kukreti R, Saso L, Kukreti S. Oxidative stress: a key modulator in neurodegenerative diseases. *Molecules*. 2019;24(8):1583. doi: [10.3390/molecules24081583](https://doi.org/10.3390/molecules24081583)
 15. Hollingworth P, Harold D, Sims R, Gerrish A, Lambert JC, Carrasquillo MM, et al. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat Genet*. 2011;43(5):429-35. doi: [10.1038/ng.803](https://doi.org/10.1038/ng.803)
 16. Newell C, Sabouny R, Hittel DS, Shutt TE, Khan A, Klein MS, et al. Mesenchymal stem cells shift mitochondrial dynamics and enhance oxidative phosphorylation in recipient cells. *Front Physiol*. 2018;9:1572. doi: [10.3389/fphys.2018.01572](https://doi.org/10.3389/fphys.2018.01572)
 17. Lee SE, Kwon D, Shin N, Kong D, Kim NG, Kim HY, et al. Accumulation of APP-CTF induces mitophagy dysfunction in the iNSCs model of Alzheimer's disease. *Cell Death Discov*. 2022;8(1):1. doi: [10.1038/s41420-021-00796-3](https://doi.org/10.1038/s41420-021-00796-3)
 18. Martín-Maestro P, Sproul A, Martínez H, Paquet D, Gerges M, Noggle S, et al. Autophagy induction by bexarotene promotes mitophagy in presenilin 1 familial Alzheimer's disease iPSC-derived neural stem cells. *Mol Neurobiol*. 2019;56(12):8220-36. doi: [10.1007/s12035-019-01665-y](https://doi.org/10.1007/s12035-019-01665-y)
 19. Ferreira JC, Campos JC, Qvit N, Qi X, Bozi LH, Bechara LR, et al. A selective inhibitor of mitofusin 1-βIIPKC association improves heart failure outcome in rats. *Nat Commun*. 2019;10(1):329. doi: [10.1038/s41467-018-08276-6](https://doi.org/10.1038/s41467-018-08276-6)
 20. Sies H, Jones DP. Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat Rev Mol Cell Biol*. 2020;21(7):363-83. doi: [10.1038/s41580-020-0230-3](https://doi.org/10.1038/s41580-020-0230-3)
 21. Llanos-González E, Henares-Chavarino Á A, Pedrero-Prieto CM, García-Carpintero S, Frontiñán-Rubio J, Sancho-Bielsa FJ, et al. Interplay between mitochondrial oxidative disorders and proteostasis in Alzheimer's disease. *Front Neurosci*. 2019;13:1444. doi: [10.3389/fnins.2019.01444](https://doi.org/10.3389/fnins.2019.01444)
 22. Cassidy L, Fernandez F, Johnson JB, Naiker M, Owoola AG, Broszczak DA. Oxidative stress in Alzheimer's disease: a review on emergent natural polyphenolic therapeutics. *Complement Ther Med*. 2020;49:102294. doi: [10.1016/j.ctim.2019.102294](https://doi.org/10.1016/j.ctim.2019.102294)
 23. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol*. 2007;39(1):44-84. doi: [10.1016/j.biocel.2006.07.001](https://doi.org/10.1016/j.biocel.2006.07.001)
 24. Zhang XD, Wang Y, Wu JC, Lin F, Han R, Han F, et al. Down-regulation of Bcl-2 enhances autophagy activation and cell death induced by mitochondrial dysfunction in rat striatum. *J Neurosci Res*. 2009;87(16):3600-10. doi: [10.1002/jnr.22152](https://doi.org/10.1002/jnr.22152)
 25. Rak M, Bénit P, Chrétien D, Bouchereau J, Schiff M, El-Khoury R, et al. Mitochondrial cytochrome c oxidase deficiency. *Clin Sci (Lond)*. 2016;130(6):393-407. doi: [10.1042/cs20150707](https://doi.org/10.1042/cs20150707)
 26. Elgenaidi IS, Spiers JP. Regulation of the phosphoprotein phosphatase 2A system and its modulation during oxidative stress: a potential therapeutic target? *Pharmacol Ther*. 2019;198:68-89. doi: [10.1016/j.pharmthera.2019.02.011](https://doi.org/10.1016/j.pharmthera.2019.02.011)
 27. Toral-Rios D, Pichardo-Rojas PS, Alonso-Vanegas M, Campos-Peña V. GSK3β and tau protein in Alzheimer's disease and epilepsy. *Front Cell Neurosci*. 2020;14:19. doi: [10.3389/fncel.2020.00019](https://doi.org/10.3389/fncel.2020.00019)
 28. Praticò D, Uryu K, Leight S, Trojanowski JQ, Lee VM. Increased lipid peroxidation precedes amyloid plaque formation in an animal model of Alzheimer amyloidosis. *J Neurosci*. 2001;21(12):4183-7. doi: [10.1523/jneurosci.21-12-04183.2001](https://doi.org/10.1523/jneurosci.21-12-04183.2001)
 29. Galbusera C, Facheris M, Magni F, Galimberti G, Sala G, Tremolada L, et al. Increased susceptibility to plasma lipid peroxidation in Alzheimer disease patients. *Curr Alzheimer Res*. 2004;1(2):103-9. doi: [10.2174/1567205043332171](https://doi.org/10.2174/1567205043332171)
 30. Butterfield DA, Reed TT, Perluigi M, De Marco C, Coccia R, Keller JN, et al. Elevated levels of 3-nitrotyrosine in brain from subjects with amnesic mild cognitive impairment: implications for the role of nitration in the progression of Alzheimer's disease. *Brain Res*. 2007;1148:243-8. doi: [10.1016/j.brainres.2007.02.084](https://doi.org/10.1016/j.brainres.2007.02.084)
 31. Gamba P, Staurengi E, Testa G, Giannelli S, Sottero B, Leonarduzzi G. A crosstalk between brain cholesterol oxidation and glucose metabolism in Alzheimer's disease. *Front Neurosci*. 2019;13:556. doi: [10.3389/fnins.2019.00556](https://doi.org/10.3389/fnins.2019.00556)
 32. Ray PD, Huang BW, Tsuji Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal*. 2012;24(5):981-90. doi: [10.1016/j.cellsig.2012.01.008](https://doi.org/10.1016/j.cellsig.2012.01.008)
 33. Swerdlow RH. Brain aging, Alzheimer's disease, and mitochondria. *Biochim Biophys Acta Mol Basis Dis*. 2011;1812(12):1630-9. doi: [10.1016/j.bbadis.2011.08.012](https://doi.org/10.1016/j.bbadis.2011.08.012)
 34. Wang X, Wang W, Li L, Perry G, Lee HG, Zhu X. Oxidative stress and mitochondrial dysfunction in Alzheimer's disease. *Biochim Biophys Acta*. 2014;1842(8):1240-7. doi: [10.1016/j.bbadis.2013.10.015](https://doi.org/10.1016/j.bbadis.2013.10.015)
 35. Tobore TO. On the central role of mitochondria dysfunction and oxidative stress in Alzheimer's disease. *Neurol Sci*. 2019;40(8):1527-40. doi: [10.1007/s10072-019-03863-x](https://doi.org/10.1007/s10072-019-03863-x)
 36. Butterfield DA, Bader Lange ML, Sultana R. Involvements of the lipid peroxidation product, HNE, in the pathogenesis and progression of Alzheimer's disease. *Biochim Biophys Acta*. 2010;1801(8):924-9. doi: [10.1016/j.bbalip.2010.02.005](https://doi.org/10.1016/j.bbalip.2010.02.005)
 37. Cheignon C, Tomas M, Bonnefont-Rousselot D, Faller P, Hureau C, Collin F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol*. 2018;14:450-64. doi: [10.1016/j.redox.2017.10.014](https://doi.org/10.1016/j.redox.2017.10.014)
 38. Ippoliti I, Ancidoni A, Da Cas R, Pierantozzi A, Vanacore N, Trotta F. Anti-dementia drugs: a descriptive study of the prescription pattern in Italy. *Neurol Sci*. 2023;44(5):1587-95. doi: [10.1007/s10072-022-06586-8](https://doi.org/10.1007/s10072-022-06586-8)
 39. Mercier C, Rollason V, Eshmaewy M, Mendes A, Frisoni GB. The treatment of behavioural and psychological symptoms in dementia: pragmatic recommendations. *Psychogeriatrics*. 2024;24(4):968-82. doi: [10.1111/psyg.13116](https://doi.org/10.1111/psyg.13116)
 40. Zhou X, Hollern D, Liao J, Andrechek E, Wang H. NMDA receptor-mediated excitotoxicity depends on the coactivation of synaptic and extrasynaptic receptors. *Cell Death Dis*. 2013;4(3):e560. doi: [10.1038/cddis.2013.82](https://doi.org/10.1038/cddis.2013.82)
 41. Calhoun A, King C, Khoury R, Grossberg GT. An evaluation of memantine ER+donepezil for the treatment of Alzheimer's disease. *Expert Opin Pharmacother*. 2018;19(15):1711-7. doi: [10.1080/14656566.2018.1519022](https://doi.org/10.1080/14656566.2018.1519022)
 42. Chowdhury S. Monoclonal antibody treatments for Alzheimer's

- disease: aducanumab and lecanemab. *Discoveries (Craiova)*. 2023;11(3):e173. doi: [10.15190/d.2023.12](https://doi.org/10.15190/d.2023.12)
43. Wang X, Sun G, Feng T, Zhang J, Huang X, Wang T, et al. Sodium oligomannate therapeutically remodels gut microbiota and suppresses gut bacterial amino acids-shaped neuroinflammation to inhibit Alzheimer's disease progression. *Cell Res*. 2019;29(10):787-803. doi: [10.1038/s41422-019-0216-x](https://doi.org/10.1038/s41422-019-0216-x)
44. Qiu J, Feng X, Chen H, Liu W, Liu W, Wu L, et al. Discovery of novel harmine derivatives as GSK-3 β /DYRK1A dual inhibitors for Alzheimer's disease treatment. *Arch Pharm (Weinheim)*. 2024;357(2):e2300404. doi: [10.1002/ardp.202300404](https://doi.org/10.1002/ardp.202300404)
45. Tonali N, Nencetti S, Orlandini E, Ciccone L. Application of PROTAC strategy to TTR-A β protein-protein interaction for the development of Alzheimer's disease drugs. *Neural Regen Res*. 2021;16(8):1554-5. doi: [10.4103/1673-5374.303017](https://doi.org/10.4103/1673-5374.303017)
46. Feldman HH, Messer K, Qiu Y, Sabbagh M, Galasko D, Turner RS, et al. Varoglutamstat: inhibiting glutamyl cyclase as a novel target of therapy in early Alzheimer's disease. *J Alzheimers Dis*. 2024;101(S1):S79-93. doi: [10.3233/jad-231126](https://doi.org/10.3233/jad-231126)
47. Zhang J, Zhang Y, Wang J, Xia Y, Zhang J, Chen L. Recent advances in Alzheimer's disease: mechanisms, clinical trials and new drug development strategies. *Signal Transduct Target Ther*. 2024;9(1):211. doi: [10.1038/s41392-024-01911-3](https://doi.org/10.1038/s41392-024-01911-3)
48. Bhuiyan NZ, Hasan MK, Mahmud Z, Hossain MS, Rahman A. Prevention of Alzheimer's disease through diet: an exploratory review. *Metabol Open*. 2023;20:100257. doi: [10.1016/j.metop.2023.100257](https://doi.org/10.1016/j.metop.2023.100257)
49. Rainey-Smith SR, Gu Y, Gardener SL, Doecke JD, Villemagne VL, Brown BM, et al. Mediterranean diet adherence and rate of cerebral A β -amyloid accumulation: data from the Australian Imaging, Biomarkers and Lifestyle Study of Ageing. *Transl Psychiatry*. 2018;8(1):238. doi: [10.1038/s41398-018-0293-5](https://doi.org/10.1038/s41398-018-0293-5)
50. Vassilaki M, Aakre JA, Syrjanen JA, Mielke MM, Geda YE, Kremers WK, et al. Mediterranean diet, its components, and amyloid imaging biomarkers. *J Alzheimers Dis*. 2018;64(1):281-90. doi: [10.3233/jad-171121](https://doi.org/10.3233/jad-171121)
51. Liu X, Yang B, Liu Q, Gao M, Luo M. The long-term neuroprotective effect of MIND and Mediterranean diet on patients with Alzheimer's disease. *Sci Rep*. 2025;15(1):32725. doi: [10.1038/s41598-025-17055-5](https://doi.org/10.1038/s41598-025-17055-5)
52. Zakrzewski W, Dobrzyński M, Szymonowicz M, Rybak Z. Stem cells: past, present, and future. *Stem Cell Res Ther*. 2019;10(1):68. doi: [10.1186/s13287-019-1165-5](https://doi.org/10.1186/s13287-019-1165-5)
53. Samsonraj RM, Raghunath M, Nurcombe V, Hui JH, van Wijnen AJ, Cool SM. Concise review: multifaceted characterization of human mesenchymal stem cells for use in regenerative medicine. *Stem Cells Transl Med*. 2017;6(12):2173-85. doi: [10.1002/sctm.17-0129](https://doi.org/10.1002/sctm.17-0129)
54. Agnello L, Ciaccio M. Neurodegenerative diseases: from molecular basis to therapy. *Int J Mol Sci*. 2022;23(21):12854. doi: [10.3390/ijms232112854](https://doi.org/10.3390/ijms232112854)
55. Yagi T, Ito D, Okada Y, Akamatsu W, Nihei Y, Yoshizaki T, et al. Modeling familial Alzheimer's disease with induced pluripotent stem cells. *Hum Mol Genet*. 2011;20(23):4530-9. doi: [10.1093/hmg/ddr394](https://doi.org/10.1093/hmg/ddr394)
56. Sarkar TJ, Quarta M, Mukherjee S, Colville A, Paine P, Doan L, et al. Transient non-integrative expression of nuclear reprogramming factors promotes multifaceted amelioration of aging in human cells. *Nat Commun*. 2020;11(1):1545. doi: [10.1038/s41467-020-15174-3](https://doi.org/10.1038/s41467-020-15174-3)
57. Deng S, Xie H, Xie B. Cell-based regenerative and rejuvenation strategies for treating neurodegenerative diseases. *Stem Cell Res Ther*. 2025;16(1):167. doi: [10.1186/s13287-025-04285-7](https://doi.org/10.1186/s13287-025-04285-7)
58. McKelvey KJ, Powell KL, Ashton AW, Morris JM, McCracken SA. Exosomes: mechanisms of uptake. *J Circ Biomark*. 2015;4:7. doi: [10.5772/61186](https://doi.org/10.5772/61186)
59. Caruso S, Poon IK. Apoptotic cell-derived extracellular vesicles: more than just debris. *Front Immunol*. 2018;9:1486. doi: [10.3389/fimmu.2018.01486](https://doi.org/10.3389/fimmu.2018.01486)
60. Borges FT, Reis LA, Schor N. Extracellular vesicles: structure, function, and potential clinical uses in renal diseases. *Braz J Med Biol Res*. 2013;46(10):824-30. doi: [10.1590/1414-431x20132964](https://doi.org/10.1590/1414-431x20132964)
61. Duarte A, Bátiz LF, Wyneken U, Lafourcade C. Potential therapies by stem cell-derived exosomes in CNS diseases: focusing on the neurogenic niche. *Stem Cells Int*. 2016;2016:5736059. doi: [10.1155/2016/5736059](https://doi.org/10.1155/2016/5736059)
62. Xin H, Li Y, Chopp M. Exosomes/miRNAs as mediating cell-based therapy of stroke. *Front Cell Neurosci*. 2014;8:377. doi: [10.3389/fncel.2014.00377](https://doi.org/10.3389/fncel.2014.00377)
63. Kowal J, Tkach M, Théry C. Biogenesis and secretion of exosomes. *Curr Opin Cell Biol*. 2014;29:116-25. doi: [10.1016/j.ceb.2014.05.004](https://doi.org/10.1016/j.ceb.2014.05.004)
64. Boulanger CM, Loyer X, Rautou PE, Amabile N. Extracellular vesicles in coronary artery disease. *Nat Rev Cardiol*. 2017;14(5):259-72. doi: [10.1038/nrcardio.2017.7](https://doi.org/10.1038/nrcardio.2017.7)
65. Ribeiro MF, Zhu H, Millard RW, Fan GC. Exosomes function in pro- and anti-angiogenesis. *Curr Angiogenes*. 2013;2(1):54-9. doi: [10.2174/22115528113020020001](https://doi.org/10.2174/22115528113020020001)
66. Théry C, Witwer KW, Aikawa E, Alcaraz MJ, Anderson JD, Andriantsitohaina R, et al. Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. *J Extracell Vesicles*. 2018;7(1):1535750. doi: [10.1080/20013078.2018.1535750](https://doi.org/10.1080/20013078.2018.1535750)
67. Da Conceicao AR, Marinatto J, Pinheiro LS, Rody T, De Felice FG. The multifaceted role of extracellular vesicles in Alzheimer's disease. *J Neurochem*. 2025;169(8):e70209. doi: [10.1111/jnc.70209](https://doi.org/10.1111/jnc.70209)
68. Li J, Peng H, Zhang W, Li M, Wang N, Peng C, et al. Enhanced nose-to-brain delivery of combined small interfering RNAs using lesion-recognizing nanoparticles for the synergistic therapy of Alzheimer's disease. *ACS Appl Mater Interfaces*. 2023;15(46):53177-88. doi: [10.1021/acsami.3c08756](https://doi.org/10.1021/acsami.3c08756)
69. Cheng L, Doecke JD, Sharples RA, Villemagne VL, Fowler CJ, Rembach A, et al. Prognostic serum miRNA biomarkers associated with Alzheimer's disease shows concordance with neuropsychological and neuroimaging assessment. *Mol Psychiatry*. 2015;20(10):1188-96. doi: [10.1038/mp.2014.127](https://doi.org/10.1038/mp.2014.127)
70. Delay C, Mandemakers W, Hébert SS. MicroRNAs in Alzheimer's disease. *Neurobiol Dis*. 2012;46(2):285-90. doi: [10.1016/j.nbd.2012.01.003](https://doi.org/10.1016/j.nbd.2012.01.003)
71. Ye Y, Gao M, Shi W, Gao Y, Li Y, Yang W, et al. The immunomodulatory effects of mesenchymal stem cell-derived extracellular vesicles in Alzheimer's disease. *Front Immunol*. 2023;14:1325530. doi: [10.3389/fimmu.2023.1325530](https://doi.org/10.3389/fimmu.2023.1325530)
72. Ising C, Venegas C, Zhang S, Scheiblich H, Schmidt SV, Vieira-Saecker A, et al. NLRP3 inflammasome activation drives tau pathology. *Nature*. 2019;575(7784):669-73. doi: [10.1038/s41586-019-1769-z](https://doi.org/10.1038/s41586-019-1769-z)
73. Liang T, Zhang Y, Wu S, Chen Q, Wang L. The role of NLRP3 inflammasome in Alzheimer's disease and potential therapeutic targets. *Front Pharmacol*. 2022;13:845185. doi: [10.3389/fphar.2022.845185](https://doi.org/10.3389/fphar.2022.845185)
74. de Jesus Gonçalves RG, Vasques JF, da Silva-Junior AJ, Gubert F, Mendez-Otero R. Mesenchymal stem cell- and extracellular vesicle-based therapies for Alzheimer's disease: progress, advantages, and challenges. *Neural Regen Res*. 2023;18(8):1645-51. doi: [10.4103/1673-5374.361546](https://doi.org/10.4103/1673-5374.361546)
75. Song N, Scholtemeijer M, Shah K. Mesenchymal stem cell immunomodulation: mechanisms and therapeutic potential. *Trends Pharmacol Sci*. 2020;41(9):653-64. doi: [10.1016/j.tips.2020.06.009](https://doi.org/10.1016/j.tips.2020.06.009)
76. Squillaro T, Peluso G, Galderisi U. Clinical trials with

- mesenchymal stem cells: an update. *Cell Transplant.* 2016;25(5):829-48. doi: [10.3727/096368915x689622](https://doi.org/10.3727/096368915x689622)
77. Ranganath SH, Levy O, Inamdar MS, Karp JM. Harnessing the mesenchymal stem cell secretome for the treatment of cardiovascular disease. *Cell Stem Cell.* 2012;10(3):244-58. doi: [10.1016/j.stem.2012.02.005](https://doi.org/10.1016/j.stem.2012.02.005)
 78. Pajarinen J, Lin T, Gibon E, Kohno Y, Maruyama M, Nathan K, et al. Mesenchymal stem cell-macrophage crosstalk and bone healing. *Biomaterials.* 2019;196:80-9. doi: [10.1016/j.biomaterials.2017.12.025](https://doi.org/10.1016/j.biomaterials.2017.12.025)
 79. Shi Y, Wang Y, Li Q, Liu K, Hou J, Shao C, et al. Immunoregulatory mechanisms of mesenchymal stem and stromal cells in inflammatory diseases. *Nat Rev Nephrol.* 2018;14(8):493-507. doi: [10.1038/s41581-018-0023-5](https://doi.org/10.1038/s41581-018-0023-5)
 80. Jasim SA, Yumashev AV, Abdelbasset WK, Margiana R, Markov A, Suksatan W, et al. Shining the light on clinical application of mesenchymal stem cell therapy in autoimmune diseases. *Stem Cell Res Ther.* 2022;13(1):101. doi: [10.1186/s13287-022-02782-7](https://doi.org/10.1186/s13287-022-02782-7)
 81. Zhang ZY, Teoh SH, Hui JH, Fisk NM, Choolani M, Chan JK. The potential of human fetal mesenchymal stem cells for off-the-shelf bone tissue engineering application. *Biomaterials.* 2012;33(9):2656-72. doi: [10.1016/j.biomaterials.2011.12.025](https://doi.org/10.1016/j.biomaterials.2011.12.025)
 82. Badillo AT, Beggs KJ, Javazon EH, Tebbets JC, Flake AW. Murine bone marrow stromal progenitor cells elicit an in vivo cellular and humoral alloimmune response. *Biol Blood Marrow Transplant.* 2007;13(4):412-22. doi: [10.1016/j.bbmt.2006.12.447](https://doi.org/10.1016/j.bbmt.2006.12.447)
 83. Fennema EM, Tchang LA, Yuan H, van Blitterswijk CA, Martin I, Scherberich A, et al. Ectopic bone formation by aggregated mesenchymal stem cells from bone marrow and adipose tissue: a comparative study. *J Tissue Eng Regen Med.* 2018;12(1):e150-8. doi: [10.1002/term.2453](https://doi.org/10.1002/term.2453)
 84. Van den Bos J, Ouamari YE, Wouters K, Cools N, Wens I. Are cell-based therapies safe and effective in the treatment of neurodegenerative diseases? A systematic review with meta-analysis. *Biomolecules.* 2022;12(2):340. doi: [10.3390/biom12020340](https://doi.org/10.3390/biom12020340)
 85. Vangsness CT Jr, Sternberg H, Harris L. Umbilical cord tissue offers the greatest number of harvestable mesenchymal stem cells for research and clinical application: a literature review of different harvest sites. *Arthroscopy.* 2015;31(9):1836-43. doi: [10.1016/j.arthro.2015.03.014](https://doi.org/10.1016/j.arthro.2015.03.014)
 86. Margiana R, Markov A, Zekiy AO, Hamza MU, Al-Dabbagh KA, Al-Zubaidi SH, et al. Clinical application of mesenchymal stem cell in regenerative medicine: a narrative review. *Stem Cell Res Ther.* 2022;13(1):366. doi: [10.1186/s13287-022-03054-0](https://doi.org/10.1186/s13287-022-03054-0)
 87. Otero-Ortega L, Gómez de Frutos MC, Laso-García F, Rodríguez-Frutos B, Medina-Gutiérrez E, López JA, et al. Exosomes promote restoration after an experimental animal model of intracerebral hemorrhage. *J Cereb Blood Flow Metab.* 2018;38(5):767-79. doi: [10.1177/0271678x17708917](https://doi.org/10.1177/0271678x17708917)
 88. Alvarez-Erviti L, Seow Y, Yin H, Betts C, Lakhali S, Wood MJ. Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. *Nat Biotechnol.* 2011;29(4):341-5. doi: [10.1038/nbt.1807](https://doi.org/10.1038/nbt.1807)
 89. Kooijmans SA, Aleza CG, Roffler SR, van Solinge WW, Vader P, Schiffelers RM. Display of GPI-anchored anti-EGFR nanobodies on extracellular vesicles promotes tumour cell targeting. *J Extracell Vesicles.* 2016;5:31053. doi: [10.3402/jev.v5.31053](https://doi.org/10.3402/jev.v5.31053)
 90. Gazdic M, Volarevic V, Arsenijevic N, Stojkovic M. Mesenchymal stem cells: a friend or foe in immune-mediated diseases. *Stem Cell Rev Rep.* 2015;11(2):280-7. doi: [10.1007/s12015-014-9583-3](https://doi.org/10.1007/s12015-014-9583-3)
 91. Harrell CR, Jankovic MG, Fellabaum C, Volarevic A, Djonov V, Arsenijevic A, et al. Molecular mechanisms responsible for anti-inflammatory and immunosuppressive effects of mesenchymal stem cell-derived factors. *Adv Exp Med Biol.* 2019;1084:187-206. doi: [10.1007/5584_2018_306](https://doi.org/10.1007/5584_2018_306)
 92. Galieva LR, James V, Mukhamedshina YO, Rizvanov AA. Therapeutic potential of extracellular vesicles for the treatment of nerve disorders. *Front Neurosci.* 2019;13:163. doi: [10.3389/fnins.2019.00163](https://doi.org/10.3389/fnins.2019.00163)
 93. Matsumoto J, Stewart T, Banks WA, Zhang J. The transport mechanism of extracellular vesicles at the blood-brain barrier. *Curr Pharm Des.* 2017;23(40):6206-14. doi: [10.2174/1381612823666170913164738](https://doi.org/10.2174/1381612823666170913164738)
 94. Kusuma GD, Barabadi M, Tan JL, Morton DA, Frith JE, Lim R. To protect and to preserve: novel preservation strategies for extracellular vesicles. *Front Pharmacol.* 2018;9:1199. doi: [10.3389/fphar.2018.01199](https://doi.org/10.3389/fphar.2018.01199)
 95. Harrell CR, Fellabaum C, Jovicic N, Djonov V, Arsenijevic N, Volarevic V. Molecular mechanisms responsible for therapeutic potential of mesenchymal stem cell-derived secretome. *Cells.* 2019;8(5):467. doi: [10.3390/cells8050467](https://doi.org/10.3390/cells8050467)
 96. Bodart-Santos V, de Carvalho LR, de Godoy MA, Batista AF, Saraiva LM, Lima LG, et al. Extracellular vesicles derived from human Wharton's jelly mesenchymal stem cells protect hippocampal neurons from oxidative stress and synapse damage induced by amyloid- β oligomers. *Stem Cell Res Ther.* 2019;10(1):332. doi: [10.1186/s13287-019-1432-5](https://doi.org/10.1186/s13287-019-1432-5)
 97. Lee M, Ban JJ, Yang S, Im W, Kim M. The exosome of adipose-derived stem cells reduces β -amyloid pathology and apoptosis of neuronal cells derived from the transgenic mouse model of Alzheimer's disease. *Brain Res.* 2018;1691:87-93. doi: [10.1016/j.brainres.2018.03.034](https://doi.org/10.1016/j.brainres.2018.03.034)
 98. Katsuda T, Tsuchiya R, Kosaka N, Yoshioka Y, Takagaki K, Oki K, et al. Human adipose tissue-derived mesenchymal stem cells secrete functional neprilysin-bound exosomes. *Sci Rep.* 2013;3:1197. doi: [10.1038/srep01197](https://doi.org/10.1038/srep01197)
 99. Elia CA, Tamborini M, Rasile M, Desiato G, Marchetti S, Swuec P, et al. Intracerebral injection of extracellular vesicles from mesenchymal stem cells exerts reduced A β plaque burden in early stages of a preclinical model of Alzheimer's disease. *Cells.* 2019;8(9):1059. doi: [10.3390/cells8091059](https://doi.org/10.3390/cells8091059)
 100. Nakano M, Kubota K, Kobayashi E, Chikenji TS, Saito Y, Konari N, et al. Bone marrow-derived mesenchymal stem cells improve cognitive impairment in an Alzheimer's disease model by increasing the expression of microRNA-146a in hippocampus. *Sci Rep.* 2020;10(1):10772. doi: [10.1038/s41598-020-67460-1](https://doi.org/10.1038/s41598-020-67460-1)
 101. Cui GH, Wu J, Mou FF, Xie WH, Wang FB, Wang QL, et al. Exosomes derived from hypoxia-preconditioned mesenchymal stromal cells ameliorate cognitive decline by rescuing synaptic dysfunction and regulating inflammatory responses in APP/PS1 mice. *FASEB J.* 2018;32(2):654-68. doi: [10.1096/fj.201700600R](https://doi.org/10.1096/fj.201700600R)
 102. Katsuda T, Oki K, Ochiya T. Potential application of extracellular vesicles of human adipose tissue-derived mesenchymal stem cells in Alzheimer's disease therapeutics. *Methods Mol Biol.* 2015;1212:171-81. doi: [10.1007/7651_2014_98](https://doi.org/10.1007/7651_2014_98)
 103. Ullah MF, Ahmad A, Bhat SH, Abu-Duhier FM, Barreto GE, Ashraf GM. Impact of sex differences and gender specificity on behavioral characteristics and pathophysiology of neurodegenerative disorders. *Neurosci Biobehav Rev.* 2019;102:95-105. doi: [10.1016/j.neubiorev.2019.04.003](https://doi.org/10.1016/j.neubiorev.2019.04.003)
 104. Gazerani P. Contribution of central and peripheral glial cells in the development and persistence of itch: therapeutic implication of glial modulation. *Neuroglia.* 2023;4(1):15-27. doi: [10.3390/neuroglia4010002](https://doi.org/10.3390/neuroglia4010002)
 105. Subhramanyam CS, Wang C, Hu Q, Dheen ST. Microglia-mediated neuroinflammation in neurodegenerative diseases. *Semin Cell Dev Biol.* 2019;94:112-20. doi: [10.1016/j.semcdb.2019.05.004](https://doi.org/10.1016/j.semcdb.2019.05.004)

106. Ding M, Shen Y, Wang P, Xie Z, Xu S, Zhu Z, et al. Exosomes isolated from human umbilical cord mesenchymal stem cells alleviate neuroinflammation and reduce amyloid-beta deposition by modulating microglial activation in Alzheimer's disease. *Neurochem Res.* 2018;43(11):2165-77. doi: [10.1007/s11064-018-2641-5](https://doi.org/10.1007/s11064-018-2641-5)
107. Barnett R. Alzheimer's disease. *Lancet.* 2019;393(10181):1589. doi: [10.1016/s0140-6736\(19\)30851-7](https://doi.org/10.1016/s0140-6736(19)30851-7)
108. Weigel M, Wang L, Fu MM. Microtubule organization and dynamics in oligodendrocytes, astrocytes, and microglia. *Dev Neurobiol.* 2021;81(3):310-20. doi: [10.1002/dneu.22753](https://doi.org/10.1002/dneu.22753)
109. Yu W, Ying J, Wang X, Liu X, Zhao T, Yoon S, et al. The involvement of lactosylceramide in central nervous system inflammation related to neurodegenerative disease. *Front Aging Neurosci.* 2021;13:691230. doi: [10.3389/fnagi.2021.691230](https://doi.org/10.3389/fnagi.2021.691230)
110. Scarfò G, Piccarducci R, Daniele S, Franzoni F, Martini C. Exploring the role of lipid-binding proteins and oxidative stress in neurodegenerative disorders: a focus on the neuroprotective effects of nutraceutical supplementation and physical exercise. *Antioxidants (Basel).* 2022;11(11):2116. doi: [10.3390/antiox11112116](https://doi.org/10.3390/antiox11112116)
111. Kwon HS, Koh SH. Neuroinflammation in neurodegenerative disorders: the roles of microglia and astrocytes. *Transl Neurodegener.* 2020;9(1):42. doi: [10.1186/s40035-020-00221-2](https://doi.org/10.1186/s40035-020-00221-2)
112. Taylor X, Cisternas P, Jury N, Martinez P, Huang X, You Y, et al. Activated endothelial cells induce a distinct type of astrocytic reactivity. *Commun Biol.* 2022;5(1):282. doi: [10.1038/s42003-022-03237-8](https://doi.org/10.1038/s42003-022-03237-8)
113. Radenovic L, Nenadic M, Ulamek-Kozioł M, Januszewski S, Czuczwar SJ, Andjus PR, et al. Heterogeneity in brain distribution of activated microglia and astrocytes in a rat ischemic model of Alzheimer's disease after 2 years of survival. *Aging (Albany NY).* 2020;12(12):12251-67. doi: [10.18632/aging.103411](https://doi.org/10.18632/aging.103411)
114. Gao Y, Qin H, Wu D, Liu C, Fang L, Wang J, et al. Walnut peptide WEKPPVSH in alleviating oxidative stress and inflammation in lipopolysaccharide-activated BV-2 microglia via the Nrf2/HO-1 and NF-κB/p38 MAPK pathways. *J Biosci Bioeng.* 2021;132(5):496-504. doi: [10.1016/j.jbiosc.2021.07.009](https://doi.org/10.1016/j.jbiosc.2021.07.009)
115. Hossain MM, Toltin AC, Gamba LM, Molina MA. Deltamethrin-evoked ER stress promotes neuroinflammation in the adult mouse hippocampus. *Cells.* 2022;11(12):1961. doi: [10.3390/cells11121961](https://doi.org/10.3390/cells11121961)
116. Marinelli S, Basilico B, Marrone MC, Ragozzino D. Microglia-neuron crosstalk: signaling mechanism and control of synaptic transmission. *Semin Cell Dev Biol.* 2019;94:138-51. doi: [10.1016/j.semcdb.2019.05.017](https://doi.org/10.1016/j.semcdb.2019.05.017)
117. Wang J, Hou Y, Zhang L, Liu M, Zhao J, Zhang Z, et al. Estrogen attenuates traumatic brain injury by inhibiting the activation of microglia and astrocyte-mediated neuroinflammatory responses. *Mol Neurobiol.* 2021;58(3):1052-61. doi: [10.1007/s12035-020-02171-2](https://doi.org/10.1007/s12035-020-02171-2)
118. Wang J, Cheng C, Liu Z, Lin Y, Yang L, Zhang Z, et al. Inhibition of A1 astrocytes and activation of A2 astrocytes for the treatment of spinal cord injury. *Neurochem Res.* 2023;48(3):767-80. doi: [10.1007/s11064-022-03820-9](https://doi.org/10.1007/s11064-022-03820-9)
119. Baror R, Neumann B, Segel M, Chalut KJ, Fancy SP, Schafer DP, et al. Transforming growth factor-beta renders ageing microglia inhibitory to oligodendrocyte generation by CNS progenitors. *Glia.* 2019;67(7):1374-84. doi: [10.1002/glia.23612](https://doi.org/10.1002/glia.23612)
120. El Waly B, Buttigieg E, Karakus C, Brustlein S, Debarbieux F. Longitudinal intravital microscopy reveals axon degeneration concomitant with inflammatory cell infiltration in an LPC model of demyelination. *Front Cell Neurosci.* 2020;14:165. doi: [10.3389/fncel.2020.00165](https://doi.org/10.3389/fncel.2020.00165)
121. Xiao Y, Zhang Y, Gao YH, Zhao ZH, He J, Gao R, et al. A targeted extracellular vesicles loaded with montelukast in the treatment of demyelinating diseases. *Biochem Biophys Res Commun.* 2022;594:31-7. doi: [10.1016/j.bbrc.2022.01.051](https://doi.org/10.1016/j.bbrc.2022.01.051)
122. Palanisamy CP, Pei J, Alugoju P, Anthikapalli NVA, Jayaraman S, Veeraraghavan VP, et al. New strategies of neurodegenerative disease treatment with extracellular vesicles (EVs) derived from mesenchymal stem cells (MSCs). *Theranostics.* 2023;13(12):4138-65. doi: [10.7150/thno.83066](https://doi.org/10.7150/thno.83066)
123. Ye X, Sun X, Starovoytov V, Cai Q. Parkin-mediated mitophagy in mutant hAPP neurons and Alzheimer's disease patient brains. *Hum Mol Genet.* 2015;24(10):2938-51. doi: [10.1093/hmg/ddv056](https://doi.org/10.1093/hmg/ddv056)
124. Atlante A, Valenti D, Latina V, Amadoro G. Dysfunction of mitochondria in Alzheimer's disease: ANT and VDAC interact with toxic proteins and aid to determine the fate of brain cells. *Int J Mol Sci.* 2022;23(14):7722. doi: [10.3390/ijms23147722](https://doi.org/10.3390/ijms23147722)
125. Xu F, Wu Y, Yang Q, Cheng Y, Xu J, Zhang Y, et al. Engineered extracellular vesicles with SHP2 high expression promote mitophagy for Alzheimer's disease treatment. *Adv Mater.* 2022;34(49):e2207107. doi: [10.1002/adma.202207107](https://doi.org/10.1002/adma.202207107)
126. Bleazard W, McCaffery JM, King EJ, Bale S, Mozdy A, Tieu Q, et al. The dynamin-related GTPase Dnm1 regulates mitochondrial fission in yeast. *Nat Cell Biol.* 1999;1(5):298-304. doi: [10.1038/13014](https://doi.org/10.1038/13014)
127. Smirnova E, Griparic L, Shurland DL, van der Bliek AM. Dynamin-related protein Drp1 is required for mitochondrial division in mammalian cells. *Mol Biol Cell.* 2001;12(8):2245-56. doi: [10.1091/mbc.12.8.2245](https://doi.org/10.1091/mbc.12.8.2245)
128. Losón OC, Song Z, Chen H, Chan DC. Fis1, Mff, MiD49, and MiD51 mediate Drp1 recruitment in mitochondrial fission. *Mol Biol Cell.* 2013;24(5):659-67. doi: [10.1091/mbc.E12-10-0721](https://doi.org/10.1091/mbc.E12-10-0721)
129. Wang X, Su B, Zheng L, Perry G, Smith MA, Zhu X. The role of abnormal mitochondrial dynamics in the pathogenesis of Alzheimer's disease. *J Neurochem.* 2009;109(Suppl 1):153-9. doi: [10.1111/j.1471-4159.2009.05867.x](https://doi.org/10.1111/j.1471-4159.2009.05867.x)
130. Hu C, Huang Y, Li L. Drp1-dependent mitochondrial fission plays critical roles in physiological and pathological progresses in mammals. *Int J Mol Sci.* 2017;18(1):144. doi: [10.3390/ijms18010144](https://doi.org/10.3390/ijms18010144)
131. Manczak M, Calkins MJ, Reddy PH. Impaired mitochondrial dynamics and abnormal interaction of amyloid beta with mitochondrial protein Drp1 in neurons from patients with Alzheimer's disease: implications for neuronal damage. *Hum Mol Genet.* 2011;20(13):2495-509. doi: [10.1093/hmg/ddr139](https://doi.org/10.1093/hmg/ddr139)
132. Sabouny R, Shutt TE. Reciprocal regulation of mitochondrial fission and fusion. *Trends Biochem Sci.* 2020;45(7):564-77. doi: [10.1016/j.tibs.2020.03.009](https://doi.org/10.1016/j.tibs.2020.03.009)
133. Samudio I, Fiegl M, McQueen T, Clise-Dwyer K, Andreeff M. The warburg effect in leukemia-stroma cocultures is mediated by mitochondrial uncoupling associated with uncoupling protein 2 activation. *Cancer Res.* 2008;68(13):5198-205. doi: [10.1158/0008-5472.Can-08-0555](https://doi.org/10.1158/0008-5472.Can-08-0555)
134. Maremanda KP, Sundar IK, Rahman I. Protective role of mesenchymal stem cells and mesenchymal stem cell-derived exosomes in cigarette smoke-induced mitochondrial dysfunction in mice. *Toxicol Appl Pharmacol.* 2019;385:114788. doi: [10.1016/j.taap.2019.114788](https://doi.org/10.1016/j.taap.2019.114788)
135. Zhao L, Hu C, Zhang P, Jiang H, Chen J. Mesenchymal stem cell therapy targeting mitochondrial dysfunction in acute kidney injury. *J Transl Med.* 2019;17(1):142. doi: [10.1186/s12967-019-1893-4](https://doi.org/10.1186/s12967-019-1893-4)
136. de Castro IP, Martins LM, Tufi R. Mitochondrial quality control and neurological disease: an emerging connection. *Expert Rev Mol Med.* 2010;12:e12. doi: [10.1017/s1462399410001456](https://doi.org/10.1017/s1462399410001456)
137. Tatsuta T, Langer T. Quality control of mitochondria:

- protection against neurodegeneration and ageing. *EMBO J.* 2008;27(2):306-14. doi: [10.1038/sj.emboj.7601972](https://doi.org/10.1038/sj.emboj.7601972)
138. Detmer SA, Chan DC. Functions and dysfunctions of mitochondrial dynamics. *Nat Rev Mol Cell Biol.* 2007;8(11):870-9. doi: [10.1038/nrm2275](https://doi.org/10.1038/nrm2275)
 139. Westermann B. Mitochondrial fusion and fission in cell life and death. *Nat Rev Mol Cell Biol.* 2010;11(12):872-84. doi: [10.1038/nrm3013](https://doi.org/10.1038/nrm3013)
 140. Youle RJ, Narendra DP. Mechanisms of mitophagy. *Nat Rev Mol Cell Biol.* 2011;12(1):9-14. doi: [10.1038/nrm3028](https://doi.org/10.1038/nrm3028)
 141. Cai Q, Tammineni P. Alterations in mitochondrial quality control in Alzheimer's disease. *Front Cell Neurosci.* 2016;10:24. doi: [10.3389/fncel.2016.00024](https://doi.org/10.3389/fncel.2016.00024)
 142. Falkevall A, Alikhani N, Bhushan S, Pavlov PF, Busch K, Johnson KA, et al. Degradation of the amyloid beta-protein by the novel mitochondrial peptidosome, PreP. *J Biol Chem.* 2006;281(39):29096-104. doi: [10.1074/jbc.M602532200](https://doi.org/10.1074/jbc.M602532200)
 143. Alikhani N, Guo L, Yan S, Du H, Pinho CM, Chen JX, et al. Decreased proteolytic activity of the mitochondrial amyloid- β degrading enzyme, PreP peptidosome, in Alzheimer's disease brain mitochondria. *J Alzheimers Dis.* 2011;27(1):75-87. doi: [10.3233/jad-2011-101716](https://doi.org/10.3233/jad-2011-101716)
 144. Gregori L, Fuchs C, Figueiredo-Pereira ME, Van Nostrand WE, Goldgaber D. Amyloid beta-protein inhibits ubiquitin-dependent protein degradation in vitro. *J Biol Chem.* 1995;270(34):19702-8. doi: [10.1074/jbc.270.34.19702](https://doi.org/10.1074/jbc.270.34.19702)
 145. Tseng BP, Green KN, Chan JL, Blurton-Jones M, LaFerla FM. A β inhibits the proteasome and enhances amyloid and tau accumulation. *Neurobiol Aging.* 2008;29(11):1607-18. doi: [10.1016/j.neurobiolaging.2007.04.014](https://doi.org/10.1016/j.neurobiolaging.2007.04.014)
 146. Teixeira PF, Glaser E. Processing peptidases in mitochondria and chloroplasts. *Biochim Biophys Acta.* 2013;1833(2):360-70. doi: [10.1016/j.bbamcr.2012.03.012](https://doi.org/10.1016/j.bbamcr.2012.03.012)
 147. Reddy PH. Amyloid beta-induced glycogen synthase kinase 3 β phosphorylated VDAC1 in Alzheimer's disease: implications for synaptic dysfunction and neuronal damage. *Biochim Biophys Acta.* 2013;1832(12):1913-21. doi: [10.1016/j.bbadis.2013.06.012](https://doi.org/10.1016/j.bbadis.2013.06.012)
 148. Anandatheerthavarada HK, Biswas G, Robin MA, Avadhani NG. Mitochondrial targeting and a novel transmembrane arrest of Alzheimer's amyloid precursor protein impairs mitochondrial function in neuronal cells. *J Cell Biol.* 2003;161(1):41-54. doi: [10.1083/jcb.200207030](https://doi.org/10.1083/jcb.200207030)
 149. Yao J, Irwin RW, Zhao L, Nilsen J, Hamilton RT, Brinton RD. Mitochondrial bioenergetic deficit precedes Alzheimer's pathology in female mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A.* 2009;106(34):14670-5. doi: [10.1073/pnas.0903563106](https://doi.org/10.1073/pnas.0903563106)
 150. Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, Wang N, et al. A β directly links A β to mitochondrial toxicity in Alzheimer's disease. *Science.* 2004;304(5669):448-52. doi: [10.1126/science.1091230](https://doi.org/10.1126/science.1091230)
 151. Dubey M, Chaudhury P, Kabiru H, Shea TB. Tau inhibits anterograde axonal transport and perturbs stability in growing axonal neurites in part by displacing kinesin cargo: neurofilaments attenuate tau-mediated neurite instability. *Cell Motil Cytoskeleton.* 2008;65(2):89-99. doi: [10.1002/cm.20243](https://doi.org/10.1002/cm.20243)
 152. Du H, Guo L, Yan S, Sosunov AA, McKhann GM, Yan SS. Early deficits in synaptic mitochondria in an Alzheimer's disease mouse model. *Proc Natl Acad Sci U S A.* 2010;107(43):18670-5. doi: [10.1073/pnas.1006586107](https://doi.org/10.1073/pnas.1006586107)
 153. Dragicevic N, Mamcarz M, Zhu Y, Buzzeo R, Tan J, Arendash GW, et al. Mitochondrial amyloid-beta levels are associated with the extent of mitochondrial dysfunction in different brain regions and the degree of cognitive impairment in Alzheimer's transgenic mice. *J Alzheimers Dis.* 2010;20 Suppl 2:S535-50. doi: [10.3233/jad-2010-100342](https://doi.org/10.3233/jad-2010-100342)
 154. Hansson Petersen CA, Alikhani N, Behbahani H, Wiehager B, Pavlov PF, Alafuzoff I, et al. The amyloid beta-peptide is imported into mitochondria via the TOM import machinery and localized to mitochondrial cristae. *Proc Natl Acad Sci U S A.* 2008;105(35):13145-50. doi: [10.1073/pnas.0806192105](https://doi.org/10.1073/pnas.0806192105)
 155. Xie Y, Zhou B, Lin MY, Wang S, Foust KD, Sheng ZH. Endolysosomal deficits augment mitochondria pathology in spinal motor neurons of asymptomatic fALS mice. *Neuron.* 2015;87(2):355-70. doi: [10.1016/j.neuron.2015.06.026](https://doi.org/10.1016/j.neuron.2015.06.026)
 156. Debosschere Y, Depuydt E, Pauwelyn G, Beerts C, Van Hecke L, Verhaert L, et al. Safety and immunomodulatory properties of equine peripheral blood-derived mesenchymal stem cells in healthy cats. *Vet Immunol Immunopathol.* 2020;227:110083. doi: [10.1016/j.vetimm.2020.110083](https://doi.org/10.1016/j.vetimm.2020.110083)
 157. Praveen Kumar L, Kandoi S, Misra R, Vijayalakshmi S, Rajagopal K, Verma RS. The mesenchymal stem cell secretome: a new paradigm towards cell-free therapeutic mode in regenerative medicine. *Cytokine Growth Factor Rev.* 2019;46:1-9. doi: [10.1016/j.cytofr.2019.04.002](https://doi.org/10.1016/j.cytofr.2019.04.002)
 158. Vatsa P, Negi R, Ansari UA, Khanna VK, Pant AB. Insights of extracellular vesicles of mesenchymal stem cells: a prospective cell-free regenerative medicine for neurodegenerative disorders. *Mol Neurobiol.* 2022;59(1):459-74. doi: [10.1007/s12035-021-02603-7](https://doi.org/10.1007/s12035-021-02603-7)
 159. Monakova A, Sagaradze G, Basalova N, Popov V, Balabanyan V, Efimenko A. Novel potency assay for MSC secretome-based treatment of idiopathic male infertility employed Leydig cells and revealed vascular endothelial growth factor as a promising potency marker. *Int J Mol Sci.* 2022;23(16):9414. doi: [10.3390/ijms23169414](https://doi.org/10.3390/ijms23169414)
 160. Yu H, Cheng J, Shi W, Ren B, Zhao F, Shi Y, et al. Bone marrow mesenchymal stem cell-derived exosomes promote tendon regeneration by facilitating the proliferation and migration of endogenous tendon stem/progenitor cells. *Acta Biomater.* 2020;106:328-41. doi: [10.1016/j.actbio.2020.01.051](https://doi.org/10.1016/j.actbio.2020.01.051)
 161. Alfaro M, Majcherczyk A, Kűes U, Ramírez L, Pisabarro AG. Glucose counteracts wood-dependent induction of lignocellulolytic enzyme secretion in monokaryon and dikaryon submerged cultures of the white-rot basidiomycete *Pleurotus ostreatus*. *Sci Rep.* 2020;10(1):12421. doi: [10.1038/s41598-020-68969-1](https://doi.org/10.1038/s41598-020-68969-1)
 162. Shen H, Chen J, Liu M, Zhao M, Hu D, Xie F, et al. Research progress of extracellular vesicles derived from mesenchymal stem cells in the treatment of neurodegenerative diseases. *Front Immunol.* 2025;16:1496304. doi: [10.3389/fimmu.2025.1496304](https://doi.org/10.3389/fimmu.2025.1496304)
 163. Nalivaeva NN, Zhuravin IA, Turner AJ. Neprilysin expression and functions in development, ageing and disease. *Mech Ageing Dev.* 2020;192:111363. doi: [10.1016/j.mad.2020.111363](https://doi.org/10.1016/j.mad.2020.111363)
 164. Ferreira JP, Albuquerque HM, Cardoso SM, Silva AM, Silva VL. Dual-target compounds for Alzheimer's disease: Natural and synthetic AChE and BACE-1 dual-inhibitors and their structure-activity relationship (SAR). *Eur J Med Chem.* 2021;221:113492. doi: [10.1016/j.ejmech.2021.113492](https://doi.org/10.1016/j.ejmech.2021.113492)
 165. Xiao G, Chen Q, Zhang X. MicroRNA-455-5p/CPEB1 pathway mediates A β -related learning and memory deficits in a mouse model of Alzheimer's disease. *Brain Res Bull.* 2021;177:282-94. doi: [10.1016/j.brainresbull.2021.10.008](https://doi.org/10.1016/j.brainresbull.2021.10.008)
 166. Sha S, Shen X, Cao Y, Qu L. Mesenchymal stem cells-derived extracellular vesicles ameliorate Alzheimer's disease in rat models via the microRNA-29c-3p/BACE1 axis and the Wnt/ β -catenin pathway. *Aging (Albany NY).* 2021;13(11):15285-306. doi: [10.18632/aging.203088](https://doi.org/10.18632/aging.203088)
 167. Parr C, Mirzaei N, Christian M, Sastre M. Activation of the Wnt/ β -catenin pathway represses the transcription of the β -amyloid precursor protein cleaving enzyme (BACE1) via binding of T-cell factor-4 to BACE1 promoter. *FASEB J.*

- 2015;29(2):623-35. doi: [10.1096/fj.14-253211](https://doi.org/10.1096/fj.14-253211)
168. Jeong H, Kim OJ, Oh SH, Lee S, Reum Lee HA, Lee KO, et al. Extracellular vesicles released from neprilysin gene-modified human umbilical cord-derived mesenchymal stem cell enhance therapeutic effects in an Alzheimer's disease animal model. *Stem Cells Int.* 2021;2021:5548630. doi: [10.1155/2021/5548630](https://doi.org/10.1155/2021/5548630)
169. Lee G, Neve RL, Kosik KS. The microtubule binding domain of tau protein. *Neuron.* 1989;2(6):1615-24. doi: [10.1016/0896-6273\(89\)90050-0](https://doi.org/10.1016/0896-6273(89)90050-0)
170. Tanaka Y, Kanai Y, Okada Y, Nonaka S, Takeda S, Harada A, et al. Targeted disruption of mouse conventional kinesin heavy chain, kif5B, results in abnormal perinuclear clustering of mitochondria. *Cell.* 1998;93(7):1147-58. doi: [10.1016/S0092-8674\(00\)81459-2](https://doi.org/10.1016/S0092-8674(00)81459-2)
171. Binder LI, Frankfurter A, Rebhun LI. The distribution of tau in the mammalian central nervous system. *J Cell Biol.* 1985;101(4):1371-8. doi: [10.1083/jcb.101.4.1371](https://doi.org/10.1083/jcb.101.4.1371)
172. Amos LA. Microtubule structure and its stabilisation. *Org Biomol Chem.* 2004;2(15):2153-60. doi: [10.1039/b403634d](https://doi.org/10.1039/b403634d)
173. Liu F, Iqbal K, Grundke-Iqbal I, Hart GW, Gong CX. O-GlcNAcylation regulates phosphorylation of tau: a mechanism involved in Alzheimer's disease. *Proc Natl Acad Sci U S A.* 2004;101(29):10804-9. doi: [10.1073/pnas.0400348101](https://doi.org/10.1073/pnas.0400348101)
174. Mazanetz MP, Fischer PM. Untangling tau hyperphosphorylation in drug design for neurodegenerative diseases. *Nat Rev Drug Discov.* 2007;6(6):464-79. doi: [10.1038/nrd2111](https://doi.org/10.1038/nrd2111)
175. Kuret J, Congdon EE, Li G, Yin H, Yu X, Zhong Q. Evaluating triggers and enhancers of tau fibrillization. *Microsc Res Tech.* 2005;67(3-4):141-55. doi: [10.1002/jemt.20187](https://doi.org/10.1002/jemt.20187)
176. Cheng Y, Bai F. The association of tau with mitochondrial dysfunction in Alzheimer's disease. *Front Neurosci.* 2018;12:163. doi: [10.3389/fnins.2018.00163](https://doi.org/10.3389/fnins.2018.00163)
177. Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G. Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. *Biochim Biophys Acta.* 2010;1802(1):2-10. doi: [10.1016/j.bbadis.2009.10.006](https://doi.org/10.1016/j.bbadis.2009.10.006)
178. Cabezas-Opazo FA, Vergara-Pulgar K, Pérez MJ, Jara C, Osorio-Fuentealba C, Quintanilla RA. Mitochondrial dysfunction contributes to the pathogenesis of Alzheimer's disease. *Oxid Med Cell Longev.* 2015;2015:509654. doi: [10.1155/2015/509654](https://doi.org/10.1155/2015/509654)
179. Wang ZX, Tan L, Yu JT. Axonal transport defects in Alzheimer's disease. *Mol Neurobiol.* 2015;51(3):1309-21. doi: [10.1007/s12035-014-8810-x](https://doi.org/10.1007/s12035-014-8810-x)
180. Li XC, Hu Y, Wang ZH, Luo Y, Zhang Y, Liu XP, et al. Human wild-type full-length tau accumulation disrupts mitochondrial dynamics and the functions via increasing mitofusins. *Sci Rep.* 2016;6:24756. doi: [10.1038/srep24756](https://doi.org/10.1038/srep24756)
181. Chen Z, Zhong C. Oxidative stress in Alzheimer's disease. *Neurosci Bull.* 2014;30(2):271-81. doi: [10.1007/s12264-013-1423-y](https://doi.org/10.1007/s12264-013-1423-y)
182. Manczak M, Reddy PH. Abnormal interaction of VDAC1 with amyloid beta and phosphorylated tau causes mitochondrial dysfunction in Alzheimer's disease. *Hum Mol Genet.* 2012;21(23):5131-46. doi: [10.1093/hmg/dds360](https://doi.org/10.1093/hmg/dds360)
183. Stamer K, Vogel R, Thies E, Mandelkow E, Mandelkow EM. Tau blocks traffic of organelles, neurofilaments, and APP vesicles in neurons and enhances oxidative stress. *J Cell Biol.* 2002;156(6):1051-63. doi: [10.1083/jcb.200108057](https://doi.org/10.1083/jcb.200108057)
184. Stoothoff W, Jones PB, Spiers-Jones TL, Joyner D, Chhabra E, Bercury K, et al. Differential effect of three-repeat and four-repeat tau on mitochondrial axonal transport. *J Neurochem.* 2009;111(2):417-27. doi: [10.1111/j.1471-4159.2009.06316.x](https://doi.org/10.1111/j.1471-4159.2009.06316.x)
185. Dixit R, Ross JL, Goldman YE, Holzbaur EL. Differential regulation of dynein and kinesin motor proteins by tau. *Science.* 2008;319(5866):1086-9. doi: [10.1126/science.1152993](https://doi.org/10.1126/science.1152993)
186. Wang X, Yang F, Chen P, Yang M, Deng Y, Zhan Z. Mesenchymal stem cell-derived extracellular vesicles in Alzheimer's disease: a novel cell-free therapeutic strategy and diagnostic biomarker. *Int J Nanomedicine.* 2025;20:14375-91. doi: [10.2147/ijn.S556625](https://doi.org/10.2147/ijn.S556625)
187. Serpente M, Fenoglio C, D'Anca M, Arcaro M, Sorrentino F, Visconte C, et al. MiRNA profiling in plasma neural-derived small extracellular vesicles from patients with Alzheimer's disease. *Cells.* 2020;9(6):1443. doi: [10.3390/cells9061443](https://doi.org/10.3390/cells9061443)
188. Zeng C, Meng X, Mai D, Xu K, Qu S. Overexpression of miR-132-3p contributes to neuronal protection in vitro and in vivo models of Alzheimer's disease. *Behav Brain Res.* 2022;417:113584. doi: [10.1016/j.bbr.2021.113584](https://doi.org/10.1016/j.bbr.2021.113584)
189. Lau P, Bossers K, Janky R, Salta E, Frigerio CS, Barbash S, et al. Alteration of the microRNA network during the progression of Alzheimer's disease. *EMBO Mol Med.* 2013;5(10):1613-34. doi: [10.1002/emmm.201201974](https://doi.org/10.1002/emmm.201201974)
190. Chen P, Wang F, Ling B, Zhu Y, Lin H, Huang J, et al. Mesenchymal stem cell-derived extracellular vesicles: emerging therapies for neurodegenerative diseases. *Int J Nanomedicine.* 2025;20:8547-65. doi: [10.2147/ijn.S526945](https://doi.org/10.2147/ijn.S526945)